

ANDOR

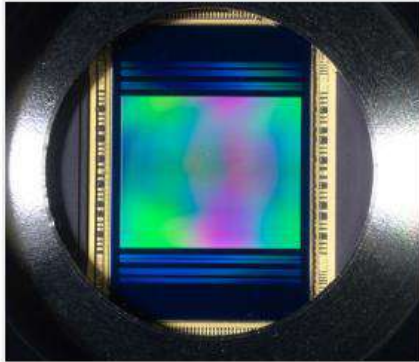
an **Oxford Instruments** company



Dragonfly
It's more than confocal...

Dragonfly High Speed Confocal:
Why Dragonfly is an ideal imaging system for you

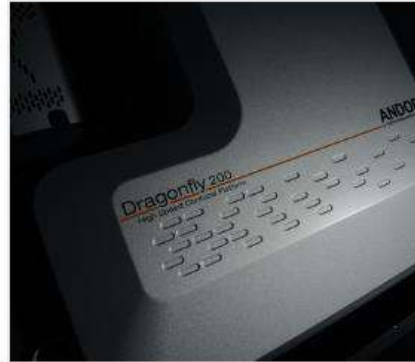
美嘉儀器客服技術部 劉思嫻
www.major.com.tw



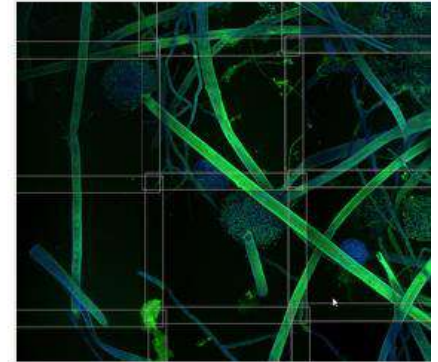
Scientific Cameras



Spectroscopy



Microscopy Systems



Software



What is an ideal (confocal) imaging system?

Fast imaging speed

- Dynamic events/large sample
- Save time

High image quality

- Keep original signals
- More details

Low photo-bleaching

- Less photo-toxicity
- More images (time/Z)



Dragonfly High Speed Confocal



Faster

20x faster than point-scanners

Deeper

3x SNR improvement in thick samples vs other multi-point scanners



Longer

10x less photo-bleaching vs point scanners

Better

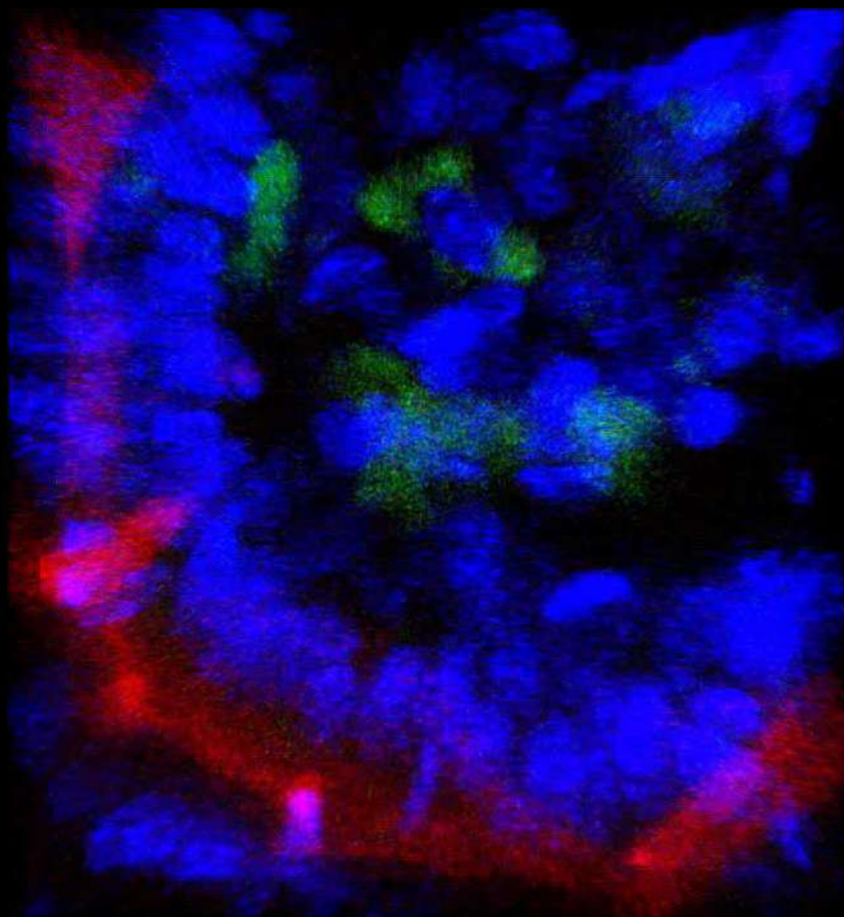
Variety of technics for better image quality (SNR/Dynamic range/Uniformity/Resolution)

Faster

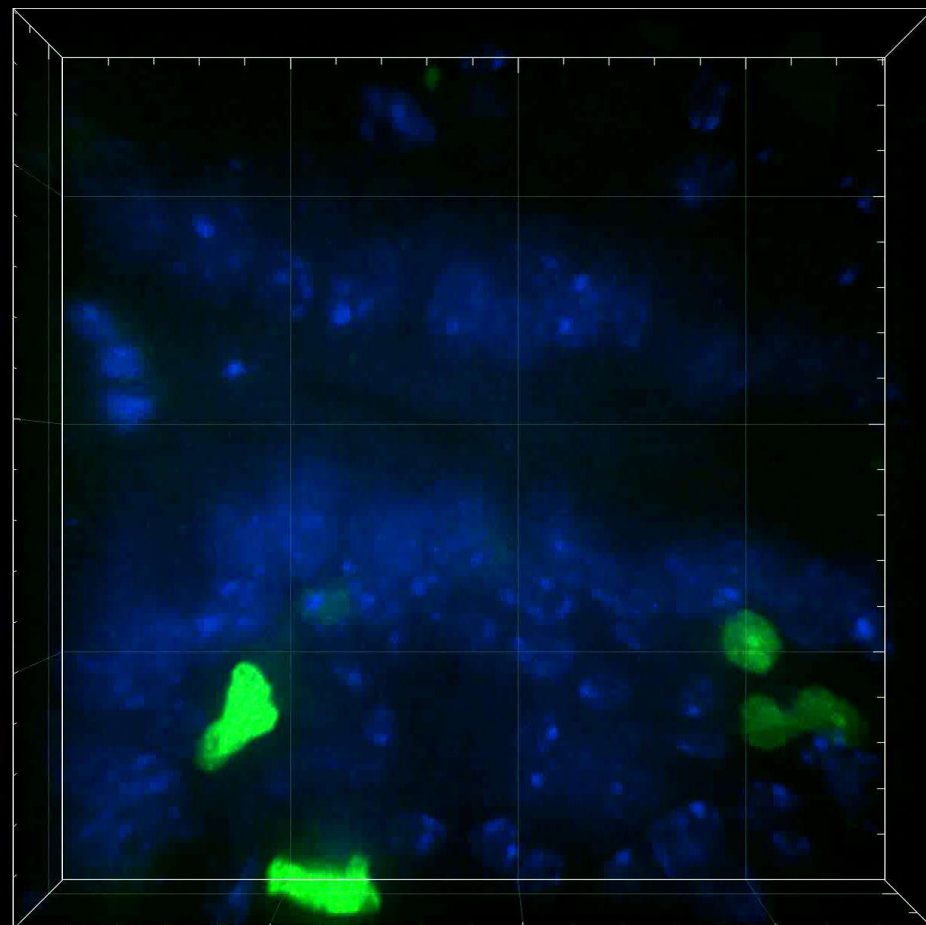
20x faster than point-scanners



Point scan confocal

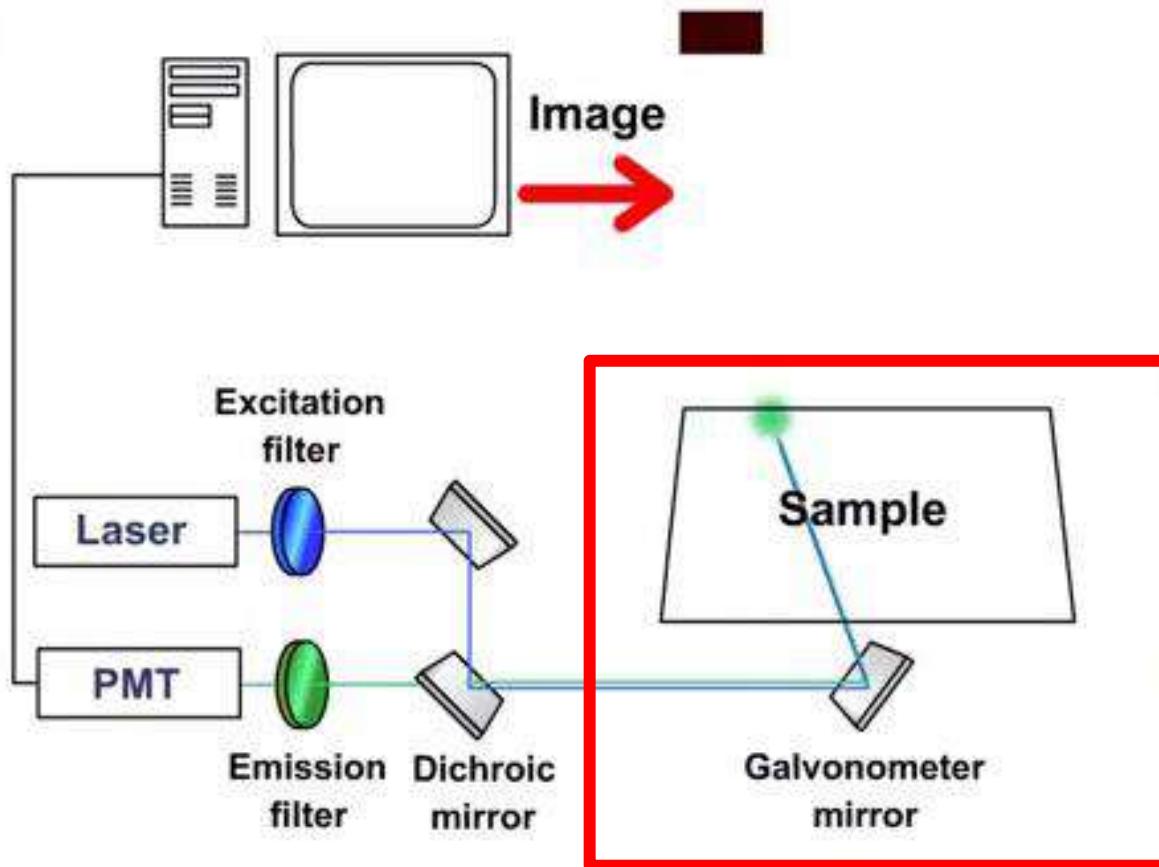


Dragonfly



Dynamic migration of Intraepithelial Lymphocytes in Mouse Intestine

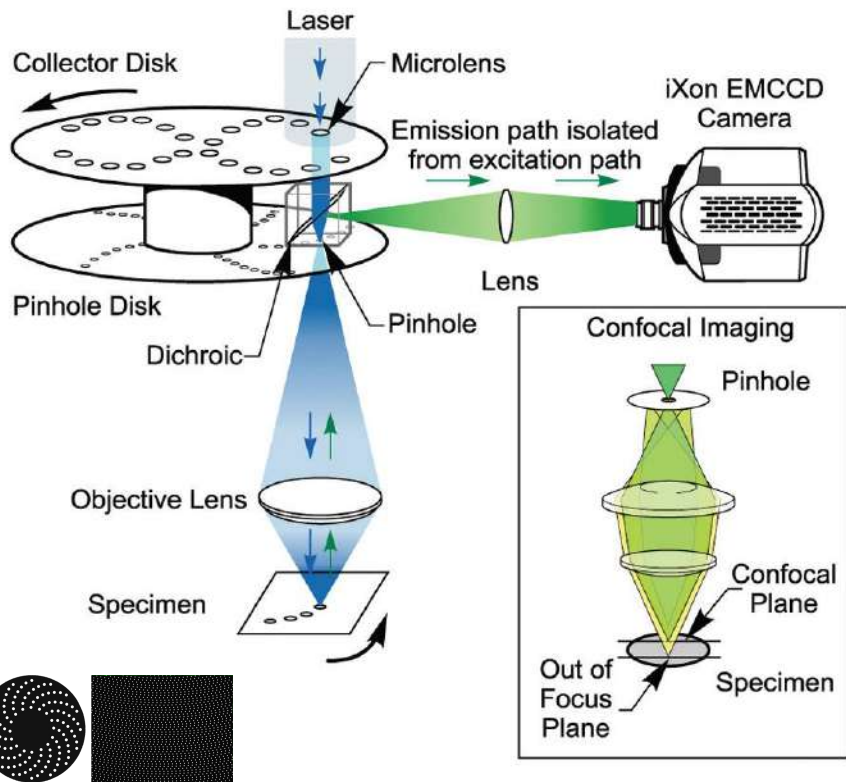
Limitation of Point-scanner



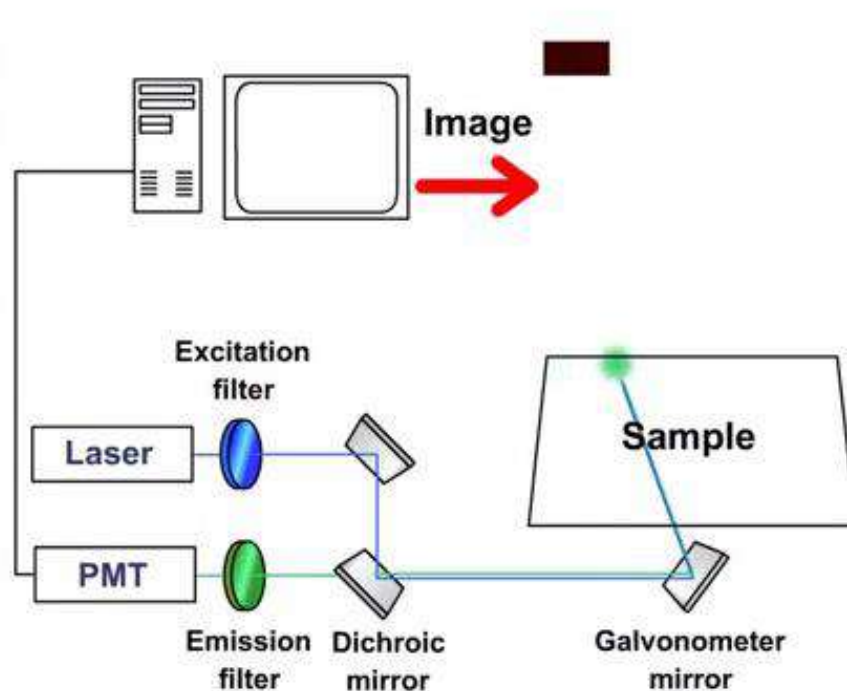
Point by point scanning

FASTER vs point-scanner

Multi-point scanning instead of single point scanning



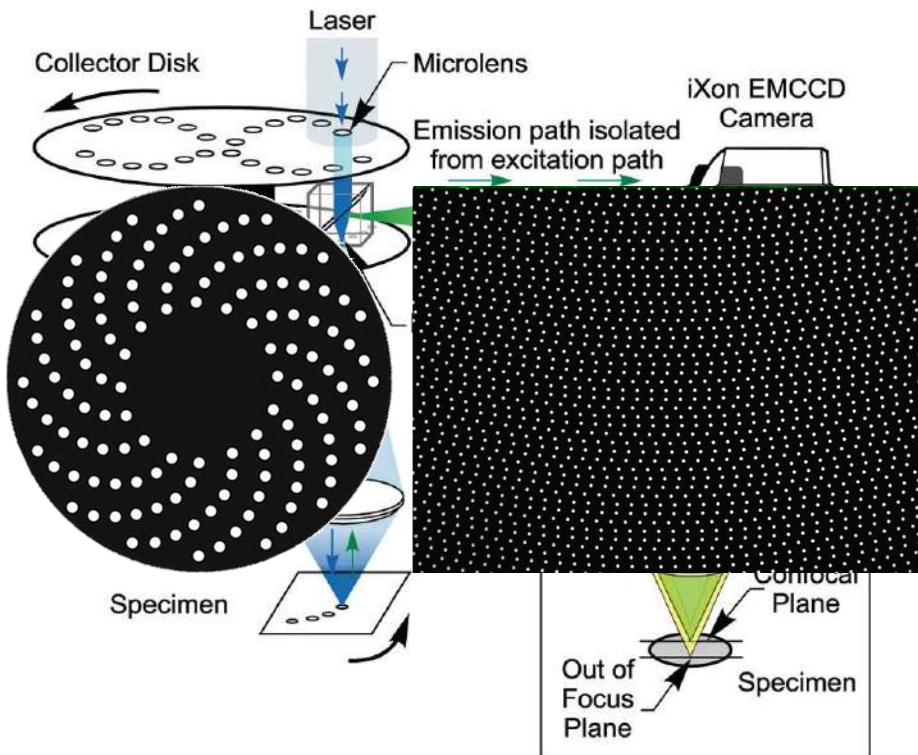
Dragonfly



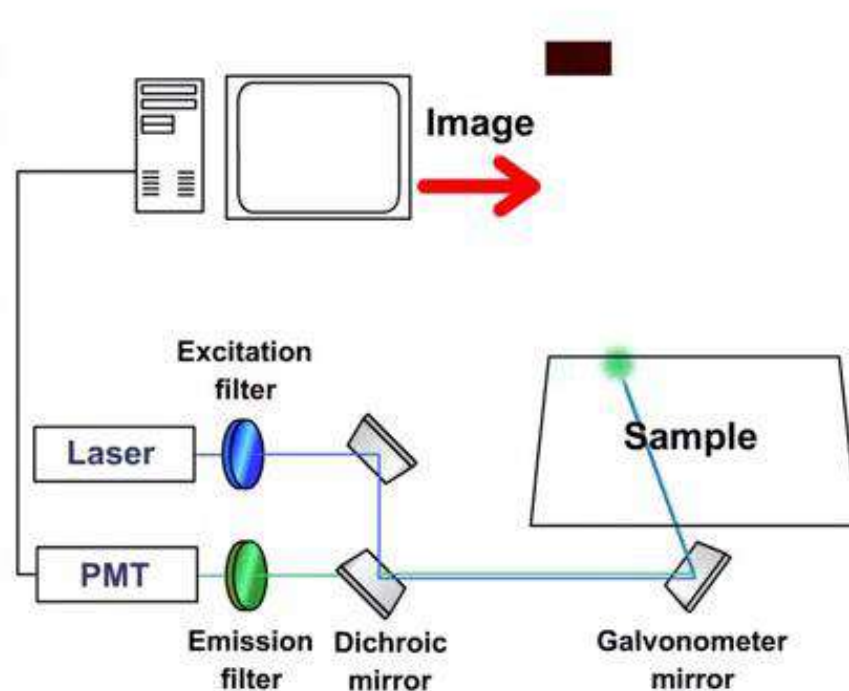
Point scanner

FASTER vs point-scanner

Multi-point scanning instead of single point scanning

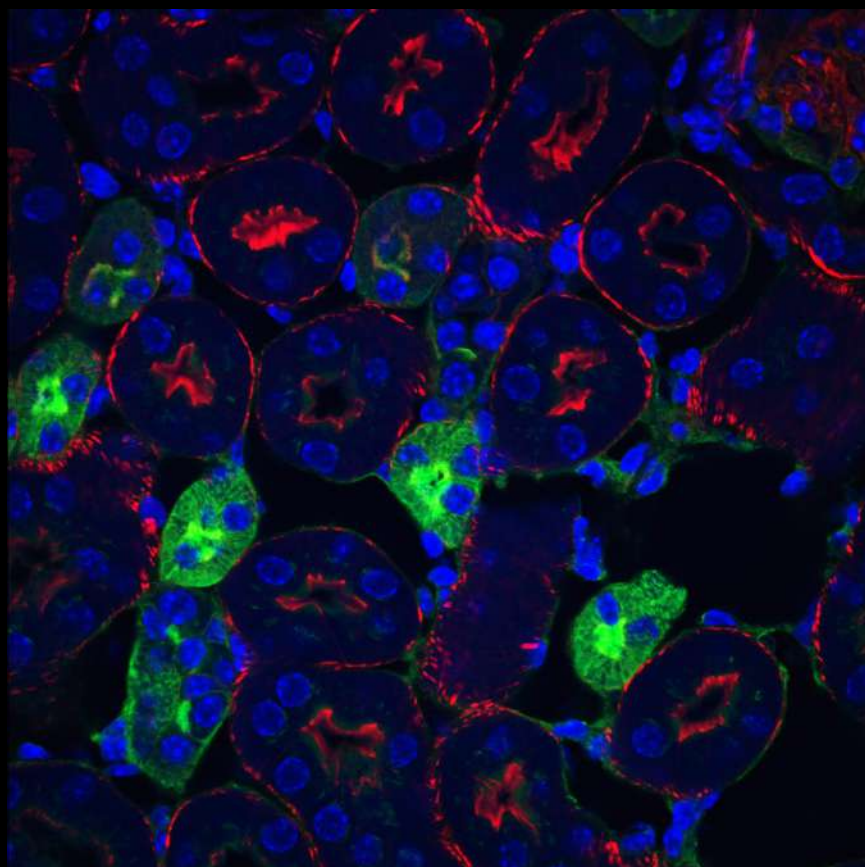


Dragonfly



Point scanner

Point scanning Confocal

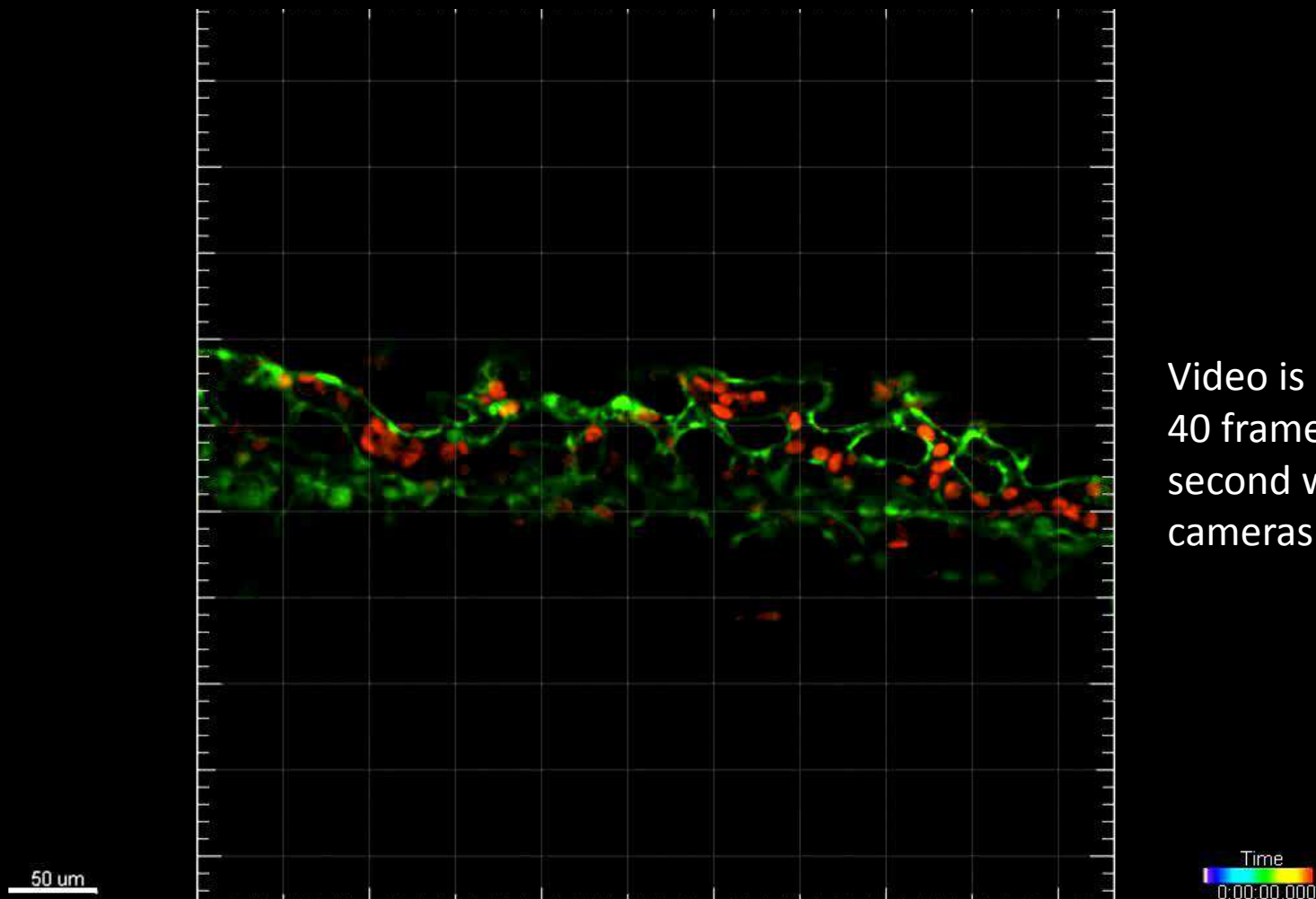


Dragonfly is an INSTANT confocal – real-time 3D display

The screenshot displays the Fusion software interface. The main window shows a dark area for the 3D display. On the right, the 'Acquisition Control' panel is visible. It includes a 'Live' button, a 'Snap' button, and a 'Channel' dropdown menu. The 'Channel' dropdown is highlighted with a yellow circle and shows 'RFP' selected. Below this, there are controls for 'Laser', 'EM Gain', and 'Exposure Time'. The 'NAVIGATION' section contains a circular control and a vertical slider. The 'SPECIMEN' section has 'New Specimen', 'Home', and 'Esc Z' buttons. The 'PROTOCOL' section shows 'Protocol 1' and 'Protocol Channels'. The 'RUN PROTOCOL' section displays 'Run Time: 00:00:27' and 'Remaining: 00:00:00'. At the bottom right, there is an 'Acquire' button.

Blood flow in transgenic zebrafish

eGFP – endothelium, DsRed - erythrocytes

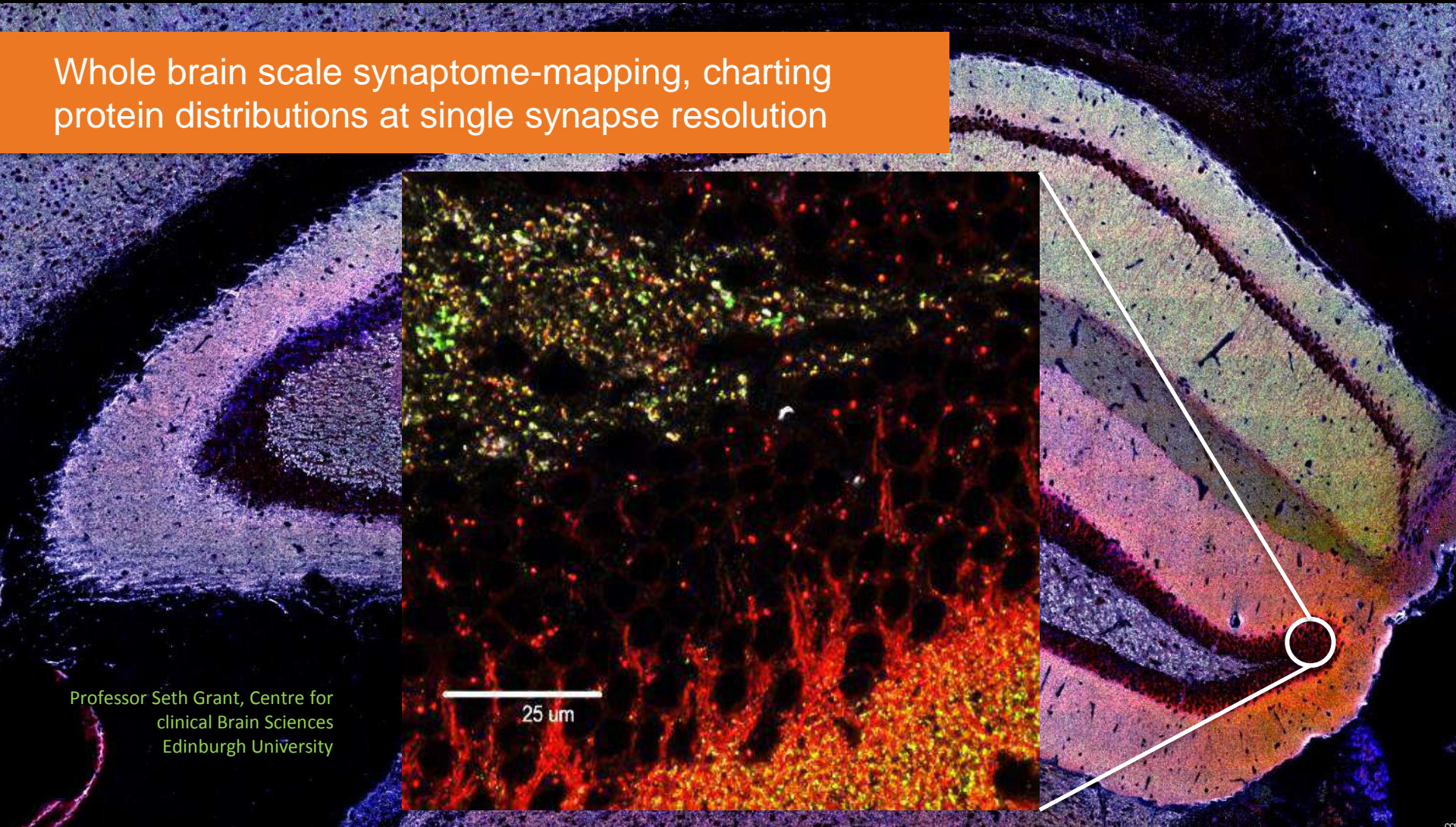


Video is real-time
40 frame-pairs per
second with two
cameras

Image courtesy of Mylène Lancino; Herbomel Lab, Institut Pasteur

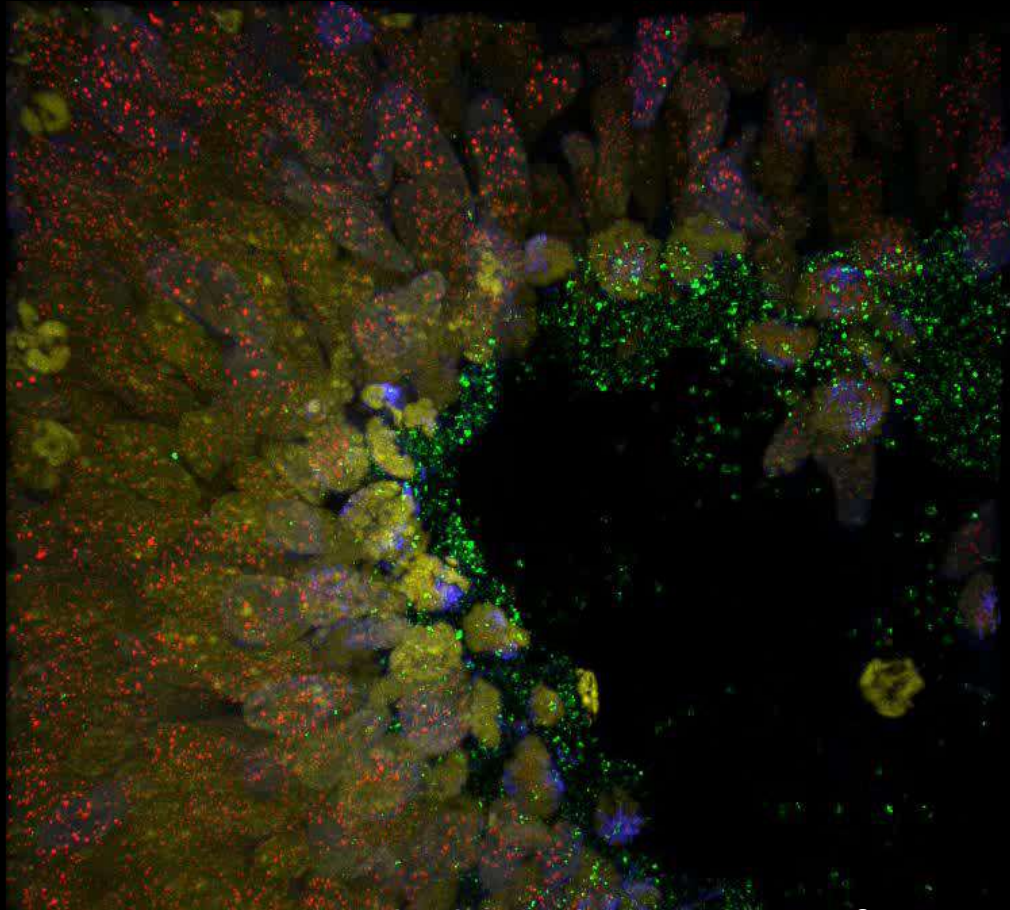
368 tile montage – 4 colours captured in 50 minutes.
Each individual image of 1024x1024 at 100x magnification.

Whole brain scale synaptome-mapping, charting protein distributions at single synapse resolution



Professor Seth Grant, Centre for
clinical Brain Sciences
Edinburgh University

Determining the stage of mitosis by the centromere number.



“Dragonfly enabled me to capture >100 high resolution images for quantification of centromere number (which are 0.2-0.8 microns in size) in at least $1/10^{\text{th}}$ of the time it would have taken on a point scanning confocal. I saved 40 hours of time in preparing a publication”

Dr Carol-Ann Martin, Andrew Jackson Lab, Institute of Genetics and Molecular Medicine, University of Edinburgh

Skin

405/488/561/640 four colors
35 Z slices per FOV, total 42 FOVs
10 mins vs >2 hrs on point scanner



Image courtesy of John CONNOLLY Lab, IMCB, A*STAR

Mouse pancreas

X – 5 (6mm)

Y – 11 (13mm)

Z – 50 (500um)

10x obj.

Imaging time

Dragonfly

40mins

Point
scanner

13hrs

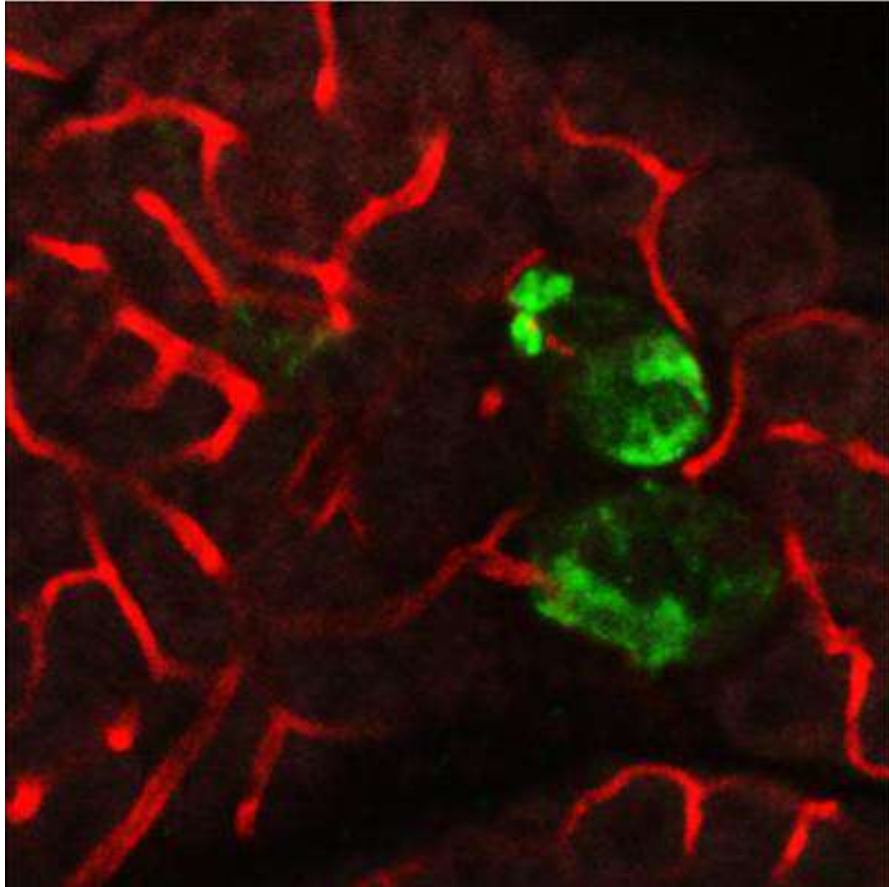
Deeper

3x SNR improvement in thick samples vs other multi-point scanners

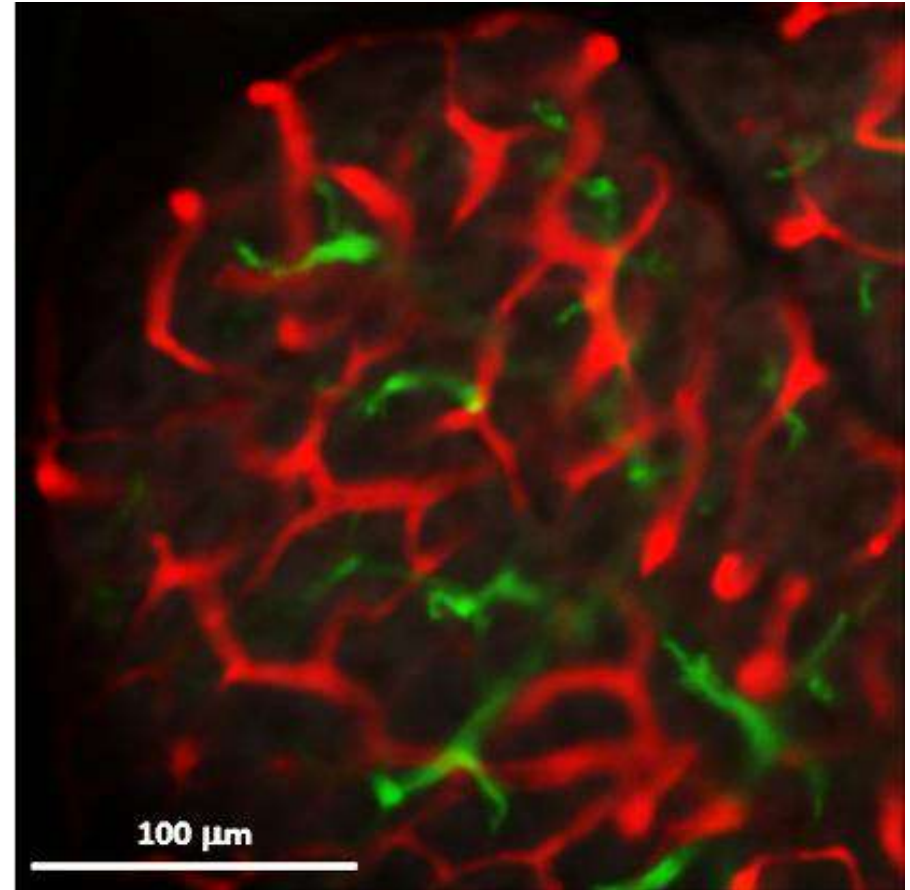


DEEPER vs other multi-point scanner

Optical section from mouse pancreas at 150um depth



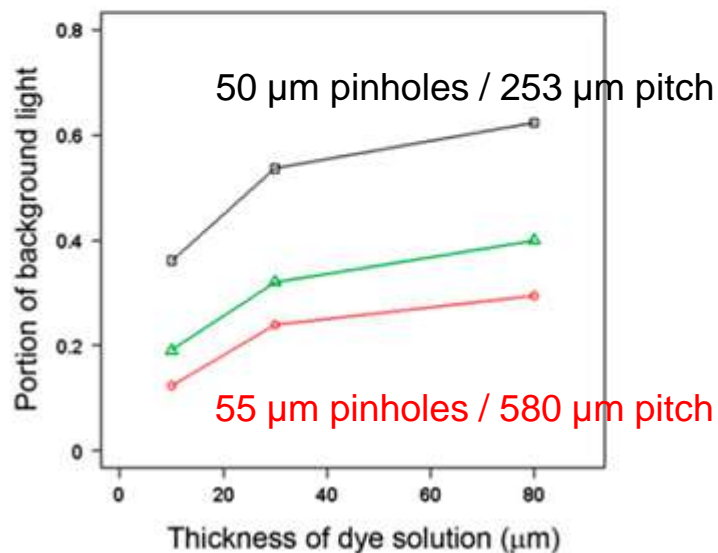
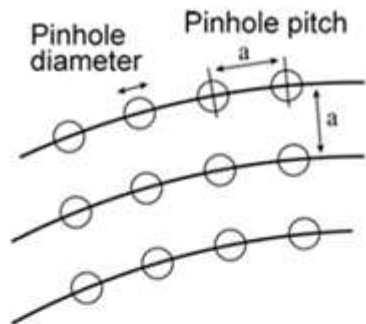
Dragonfly



Yokogawa CSU-X

DEEPER vs other multi-point scanner

Traditionally multi-beam starts to suffer with samples over 30um thickness



Traditional multi-point

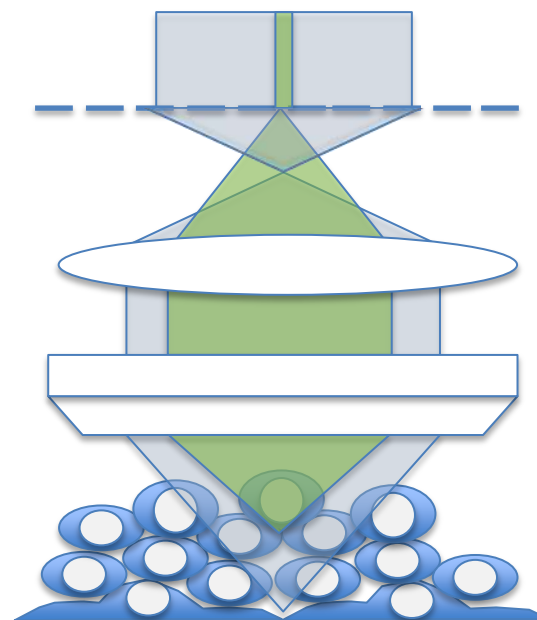
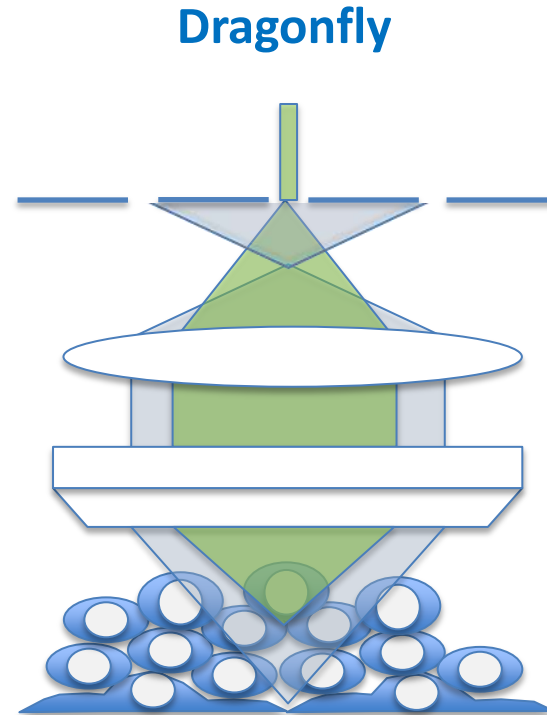
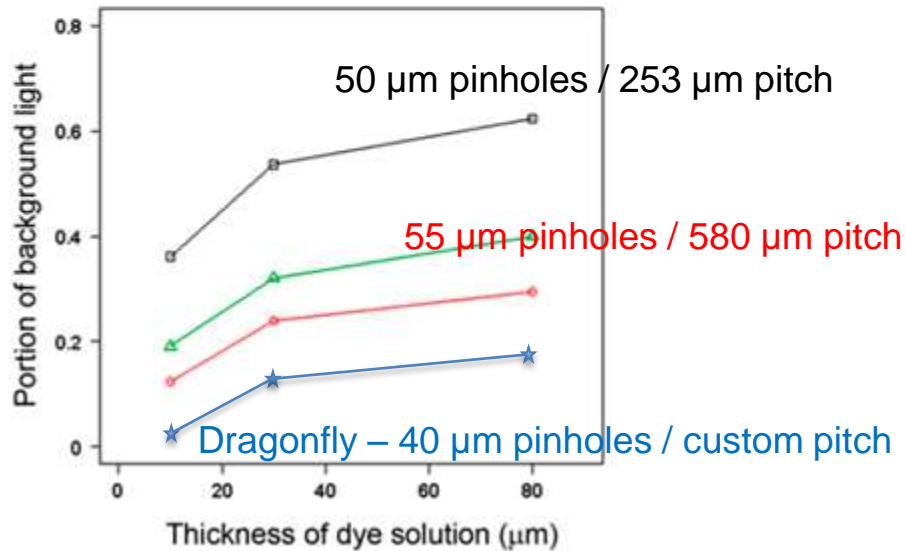
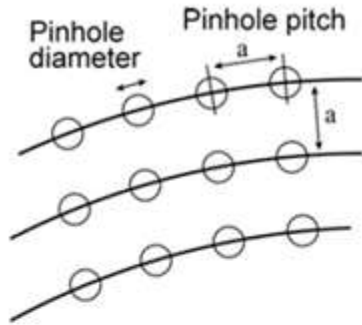


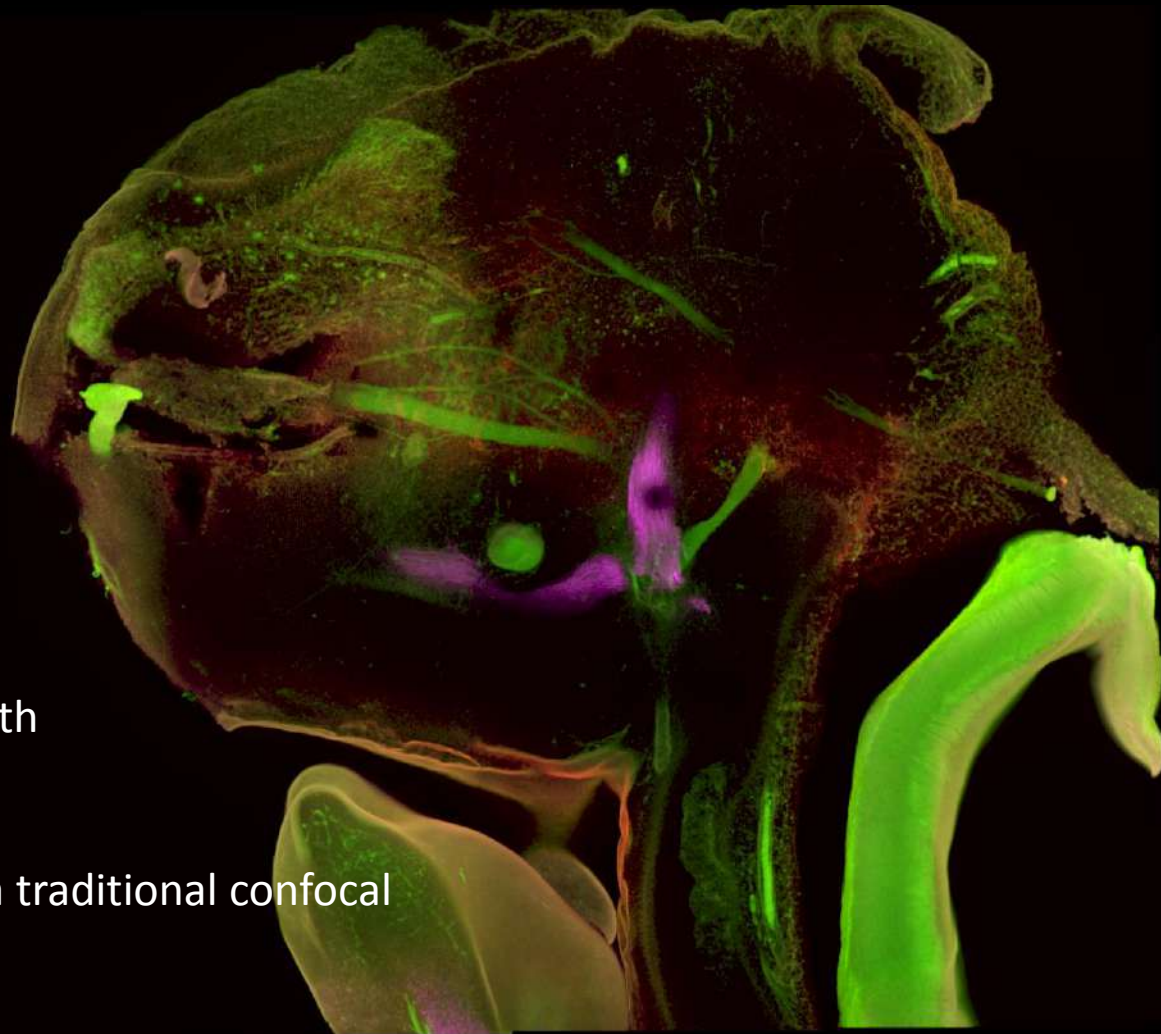
Image adapted from "Improving Spinning disk confocal microscopy by preventing pinhole cross-talk for intravital imaging" PNAS Feb. 2013; vol. 110

DEEPER vs other multi-point scanner

On Dragonfly the limiting factor in deep specimens is the transparency and refractive effects of the specimen itself



Chick Embryo Head (HH28)



10x
Tiled 7x5
Imaged to 1.2mm depth
220 Z planes
3 colours.
1hrs (vs 15 hours on a traditional confocal)

500 μm

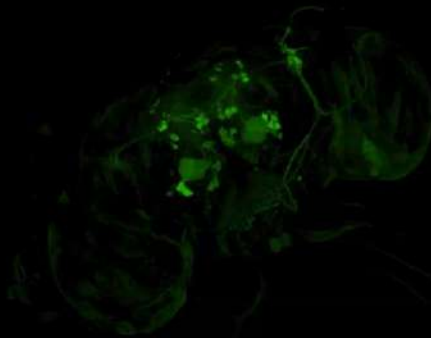
Drosophila Brain Neuron

40x water

Tiled 3x2

600 Z planes total 200um

15mins (vs 2hrs on point scanner with less Z slices)



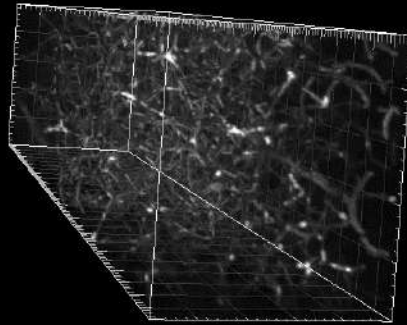
50 um

Image courtesy of Brain Research Center, NTHU, Taiwan

Cleared brain tissue

2.4 mm deep
3DISCO cleared
Perfused with red beads
8000 optical sections
20x/0.95 WI objective

400 μ m



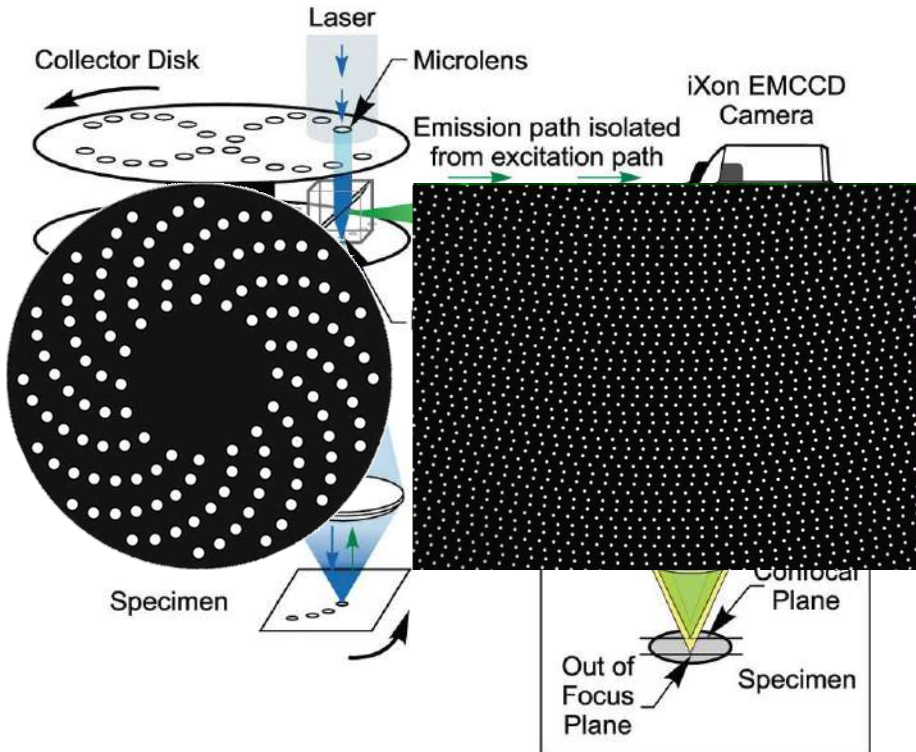
Courtesy Alan Watson, Uni Pittsburgh

Longer

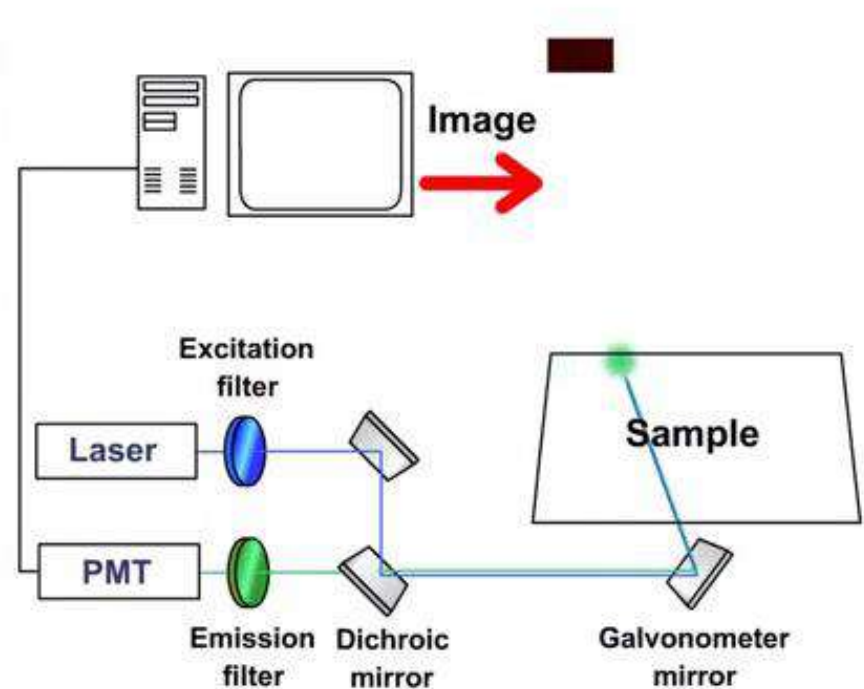
10x less photo-bleaching vs
point scanners



Multi-point scanning instead of single point scanning



Dragonfly



Point scanner

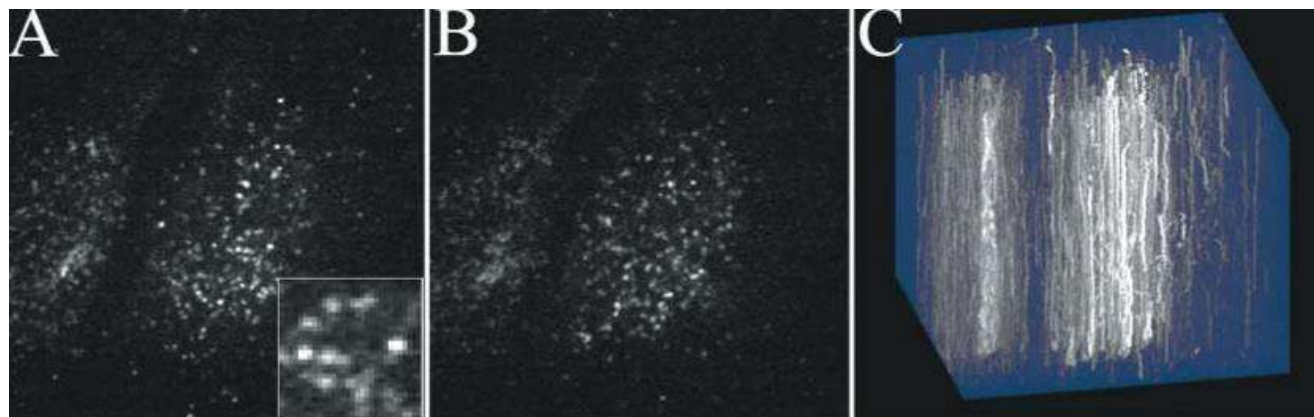
DEEPER & LONGER vs point-scanner

Low photo-bleaching for longer timelapse and more Z slices

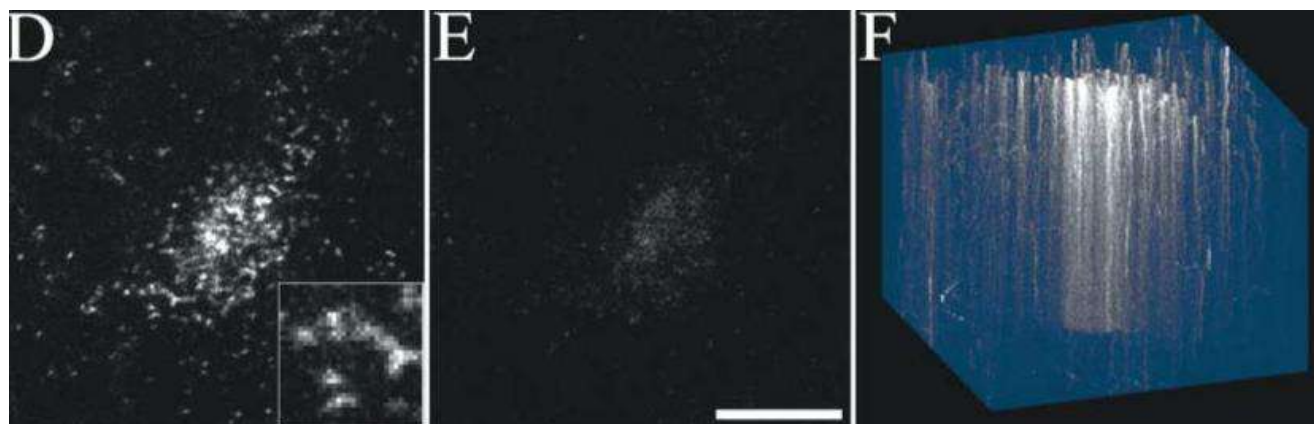
1st timepoint

200st timepoint

XY-T



Multi-point scanner

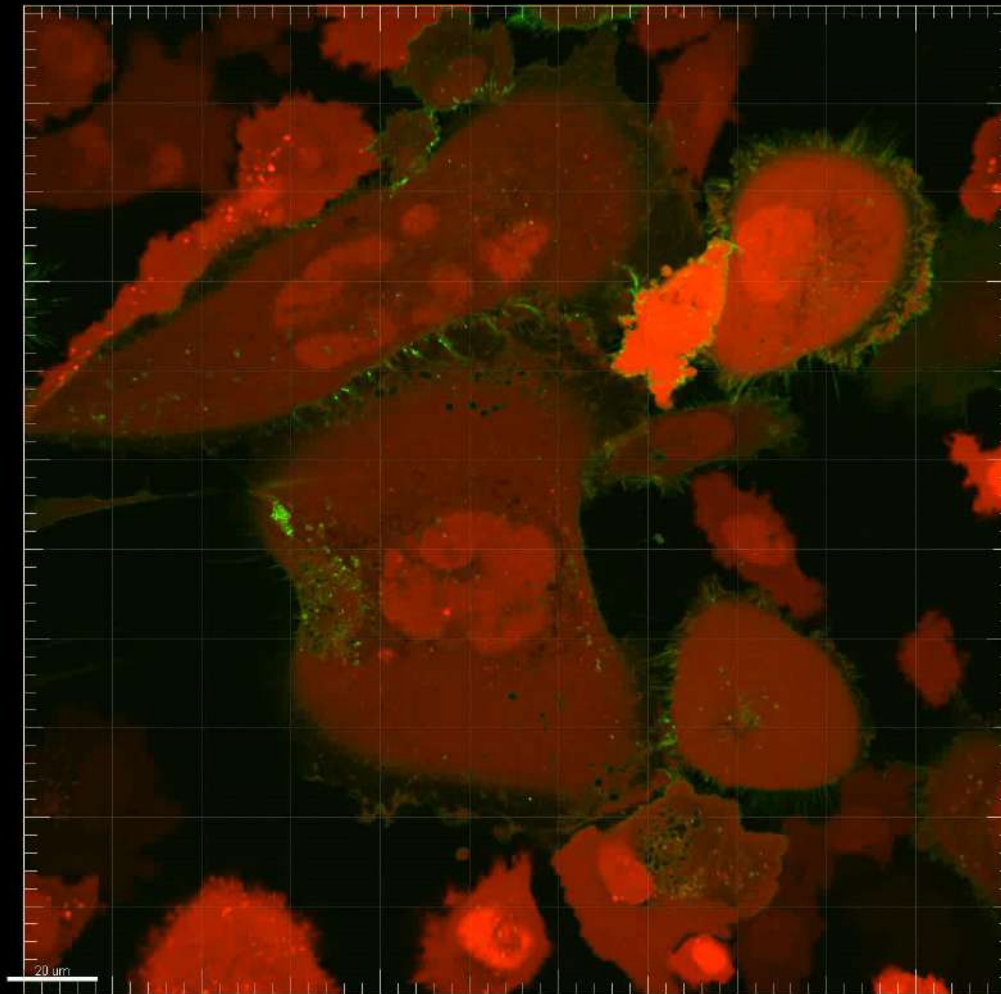


Point scanner

Data from Wang et al, Journal of Microscopy, May 2005

Live Cell Imaging overnight of Z stacks. MIP displayed.

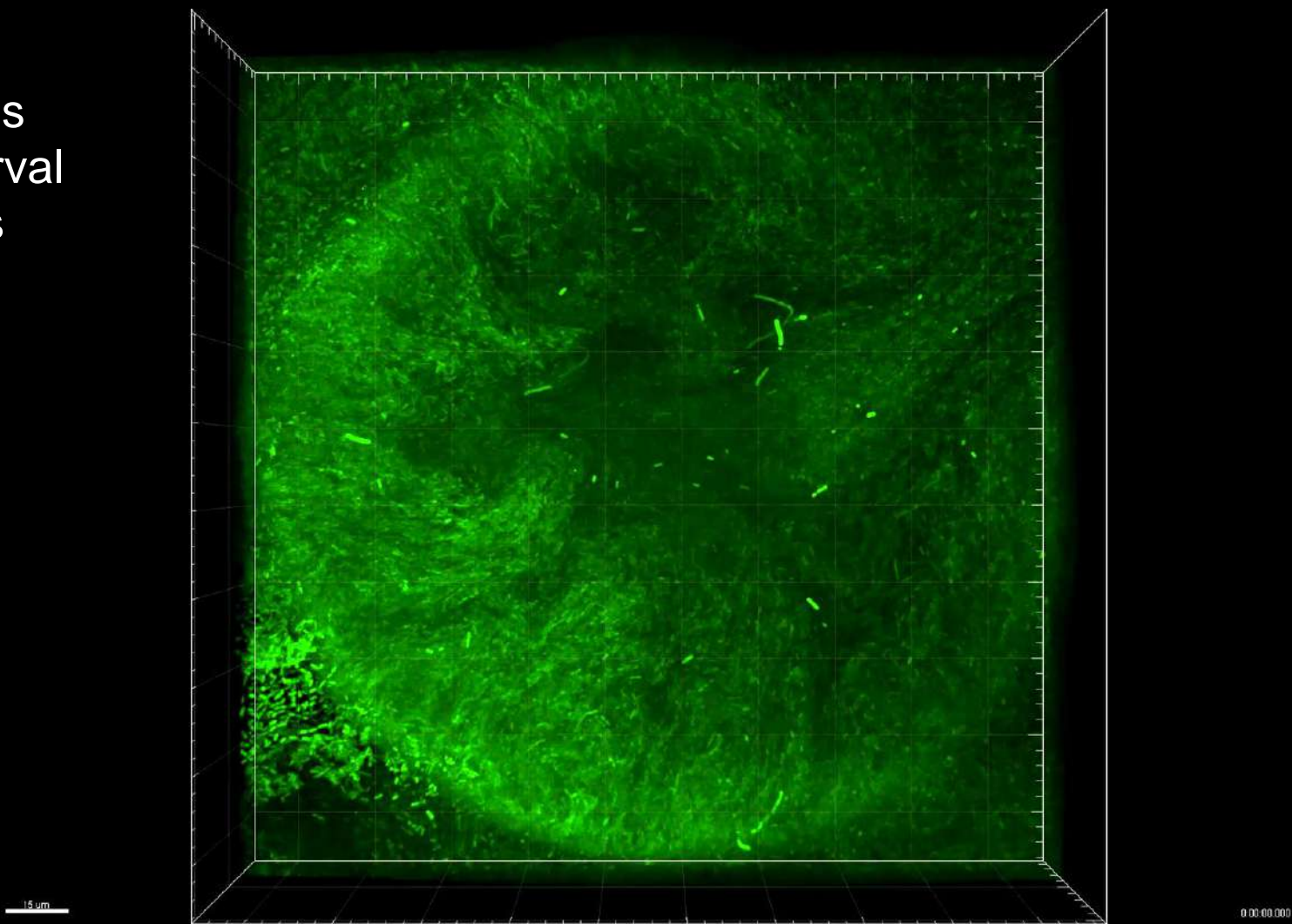
Prostate Cancer Cells. Green: EphB4GFP and Red: SUMO1mKO2



Queensland University of Technology

E.coli

60x oil
101 Z slices
4mins interval
Total 10hrs



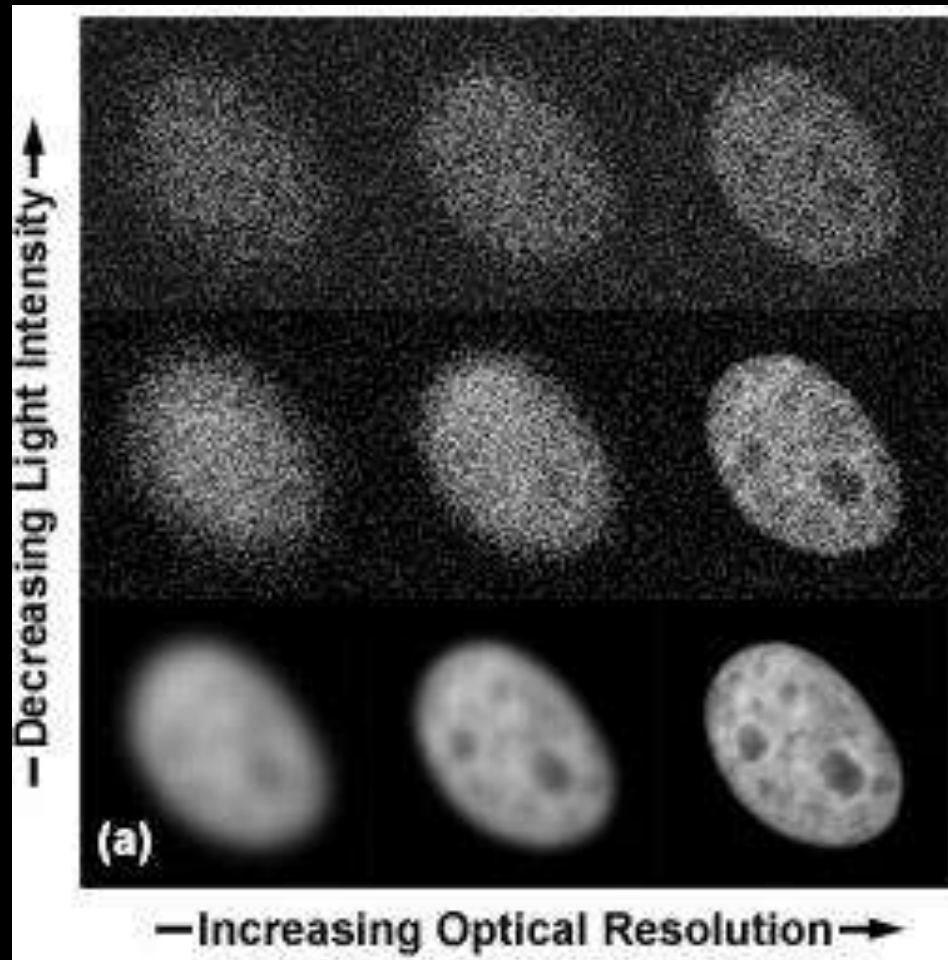
Institute of Physics, Academia Sinica, TW



Better

Variety of technics for better image quality (SNR/Dynamic range/Uniformity/Resolution)

The Importance of SNR



Photobleach, phototoxicity and sample motion all influence SNR and hence resolution

Image Quality (SNR)

Camera is 2x more sensitive than PMT

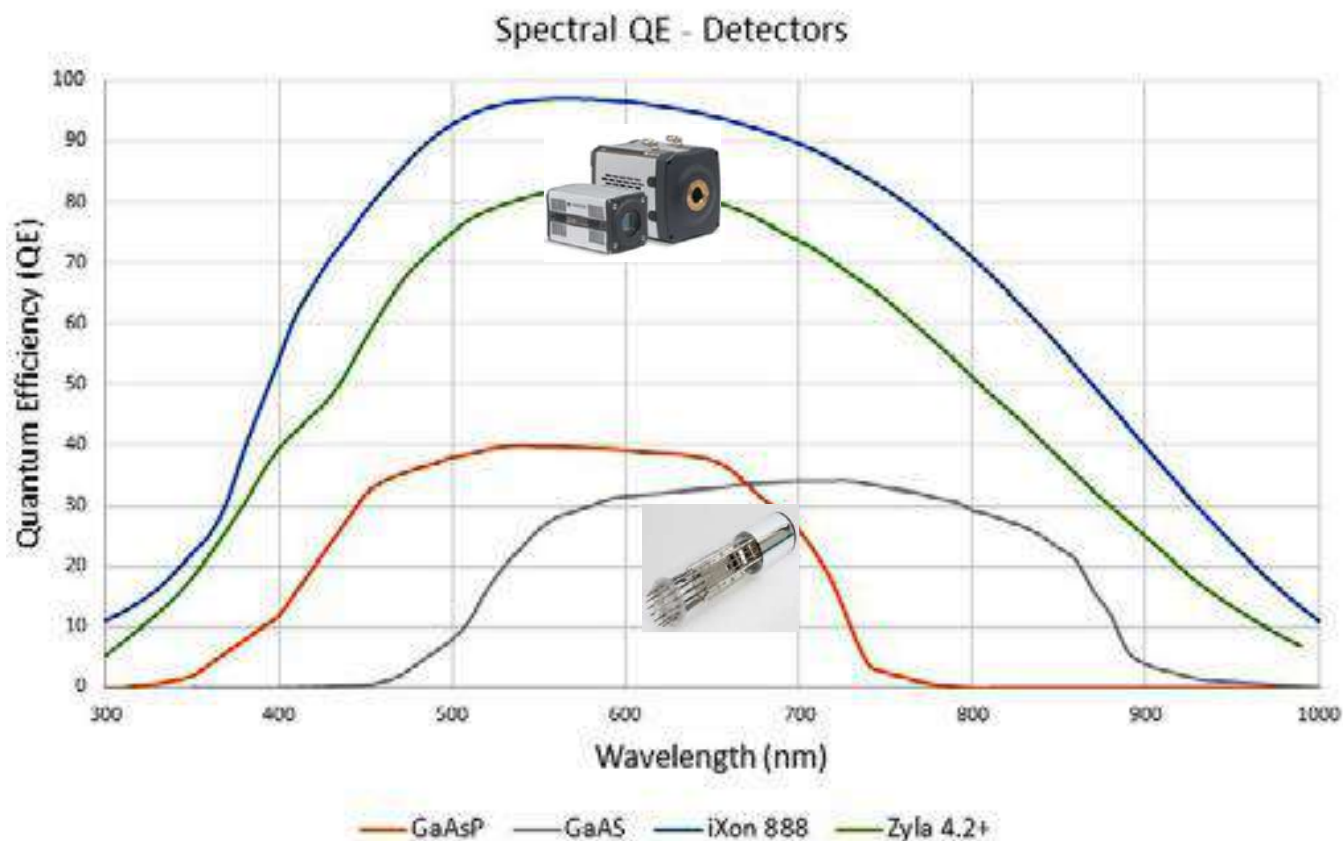


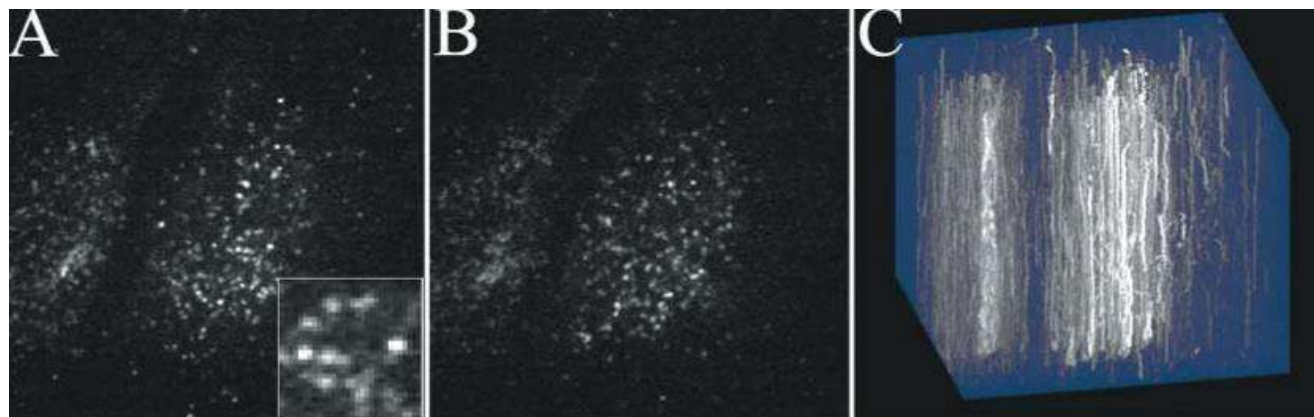
Image Quality (SNR)

Low photo-bleaching rate to keep high SNR

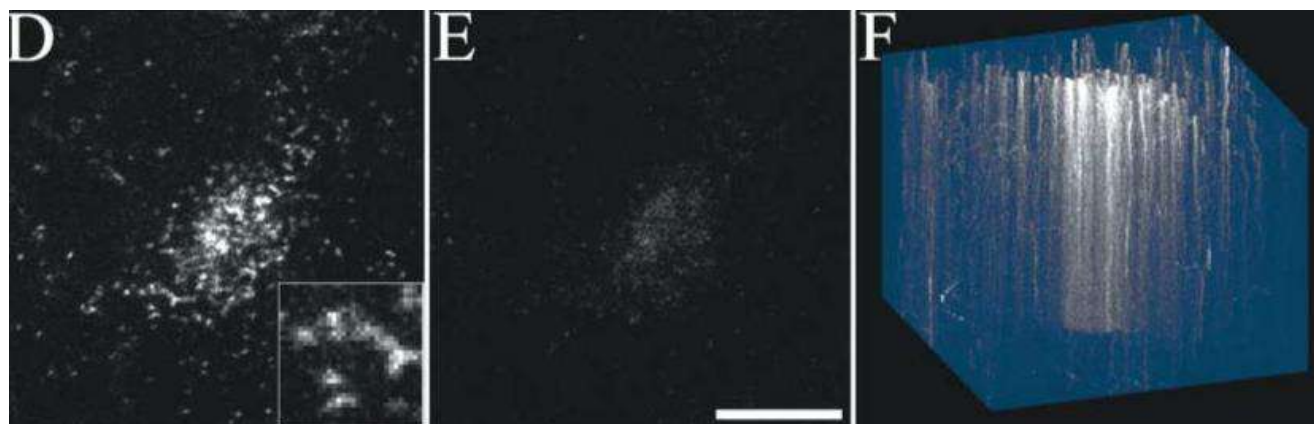
1st timepoint

200st timepoint

XY-T



Multi-point scanning



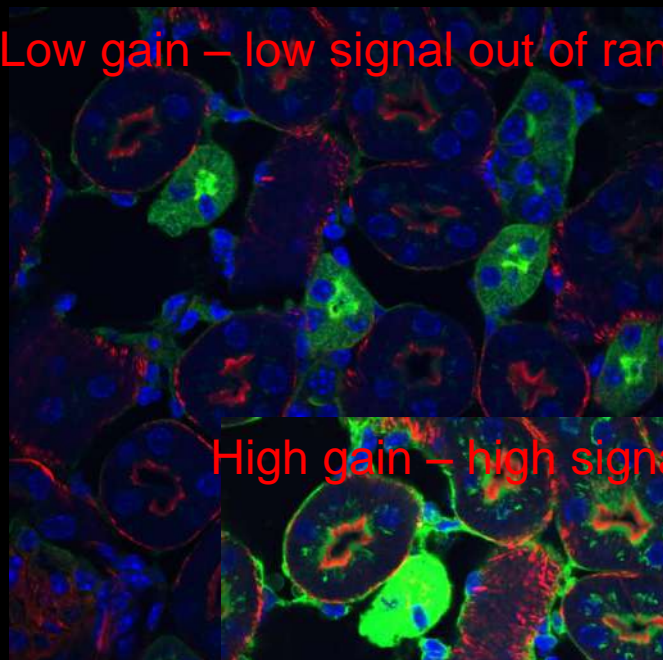
Point scanner

Data from Wang et al, Journal of Microscopy, May 2005

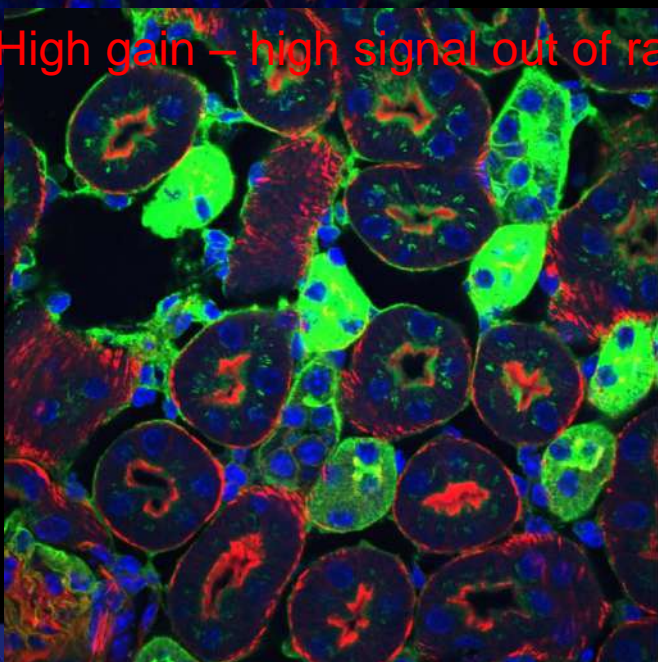
Dynamic Range

PMT-based Confocal

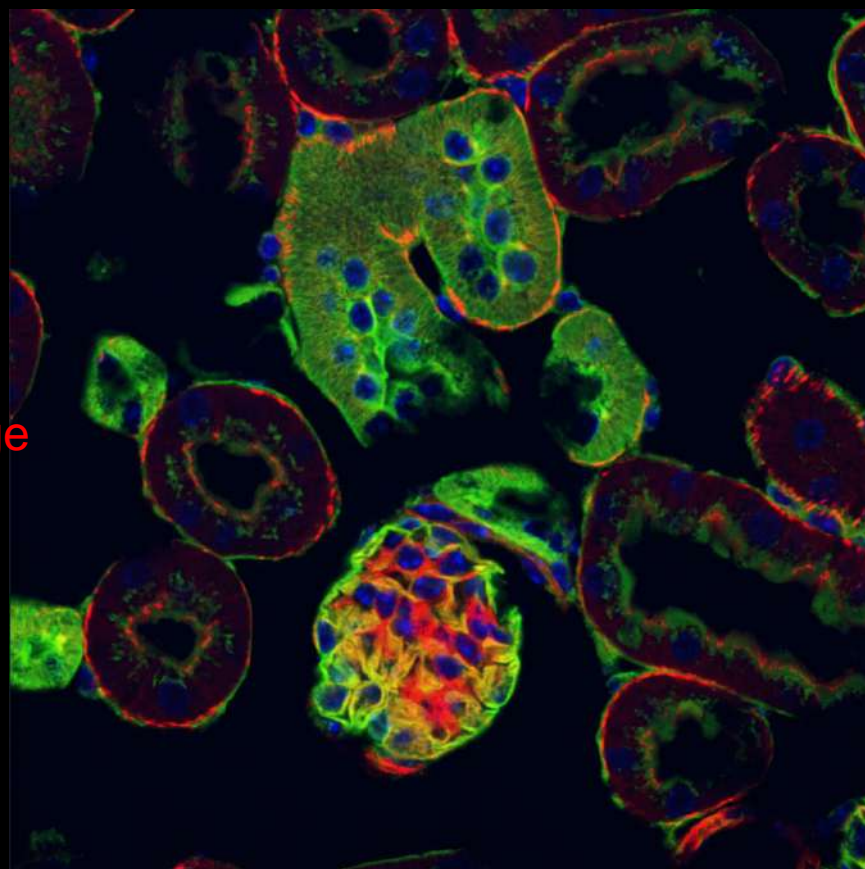
Low gain – low signal out of range



High gain – high signal out of range



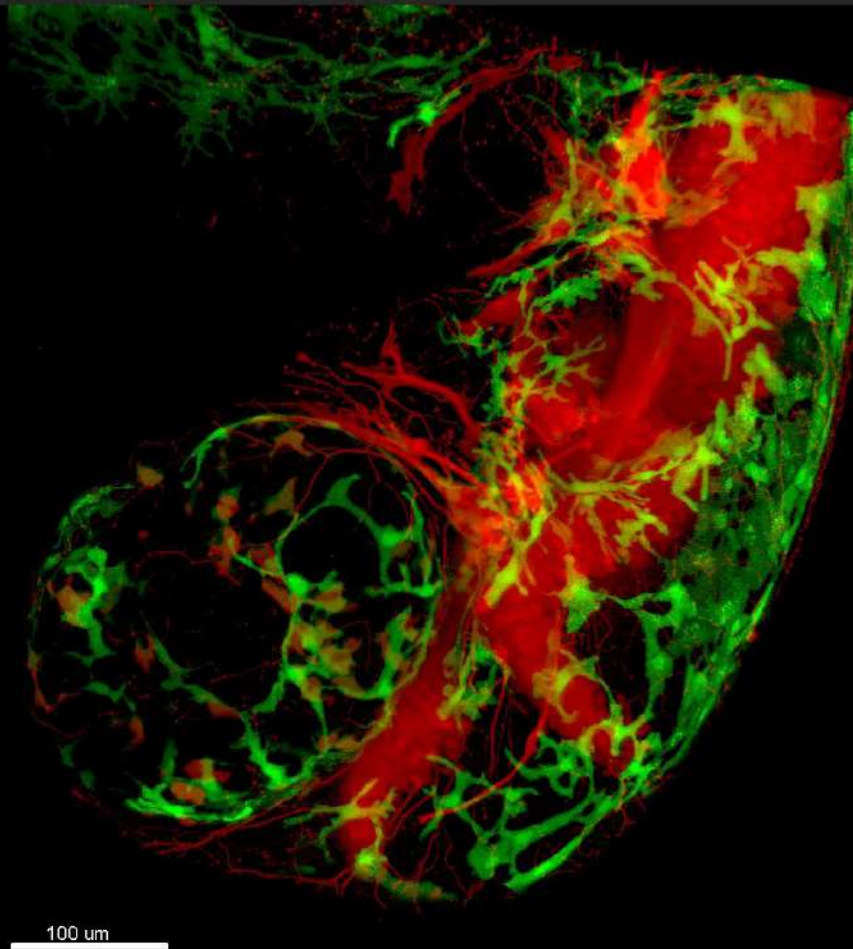
Camera-based Dragonfly Confocal



High *and* low signal in range

Zebrafish with red neural tube staining and MITFA-GFP cytoplasmic stain for melanocytes.

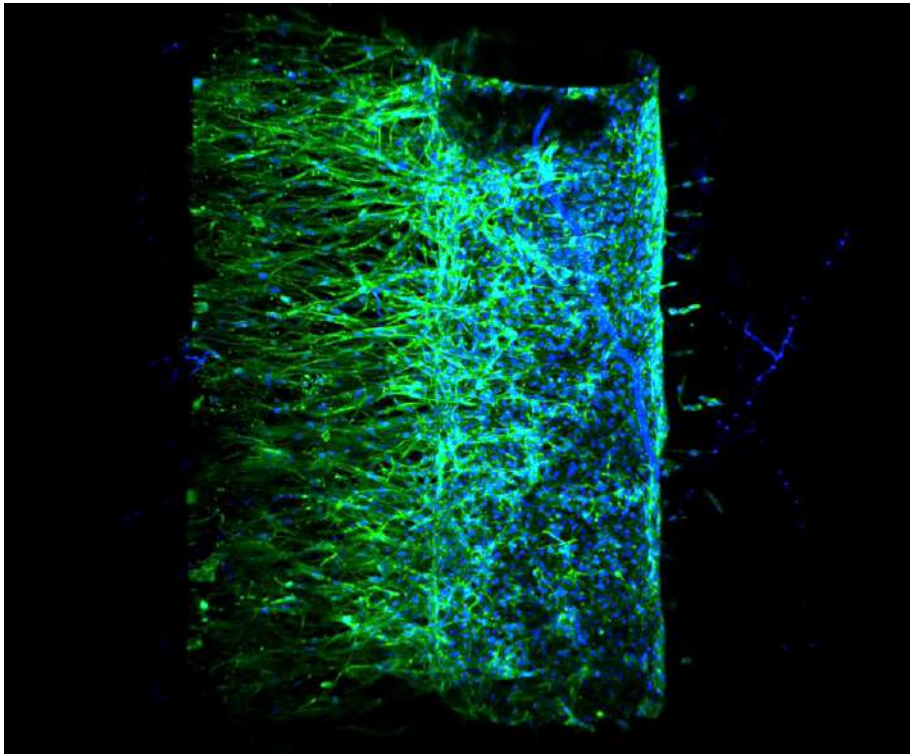
331 slices captured over 110um depth at 25x magnification with 25um pinhole



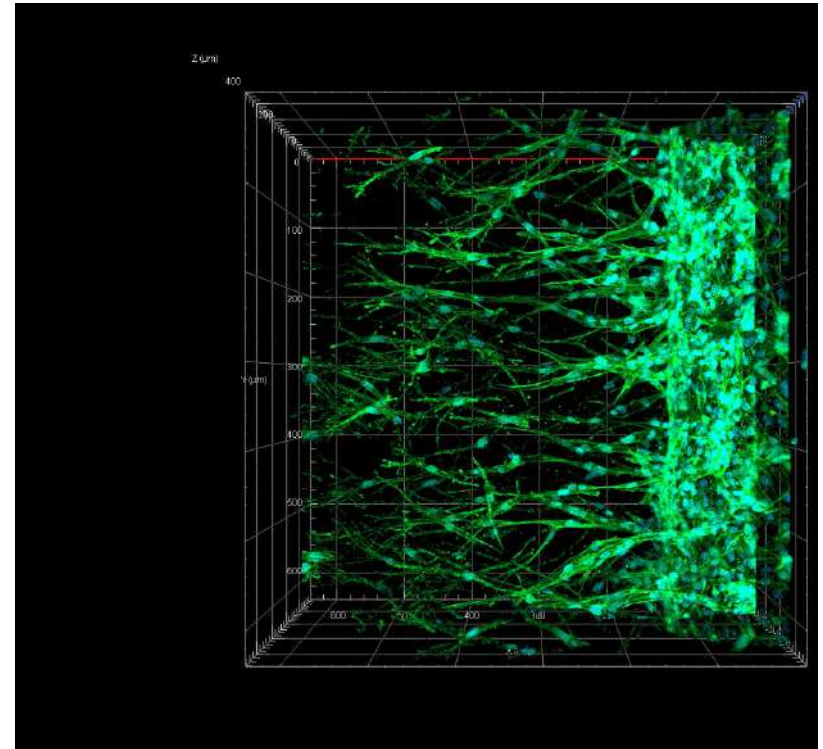
Alessandro Brombin (Patton Group), University of Edinburgh.

Image Quality (Dynamic Range)

Higher dynamic range means more details



Dragonfly



Point-scanner

Images appear “grainy or binary”;
frame averaging often required; details missed

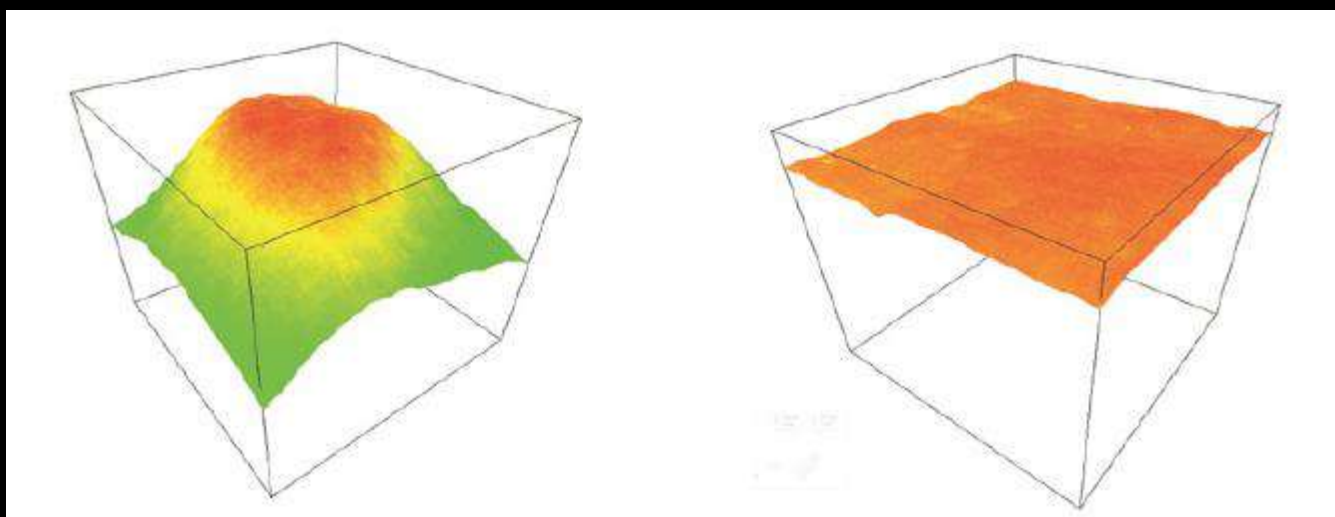
Borealis Perfect Illumination Delivery

Broad Spectrum Uniformity



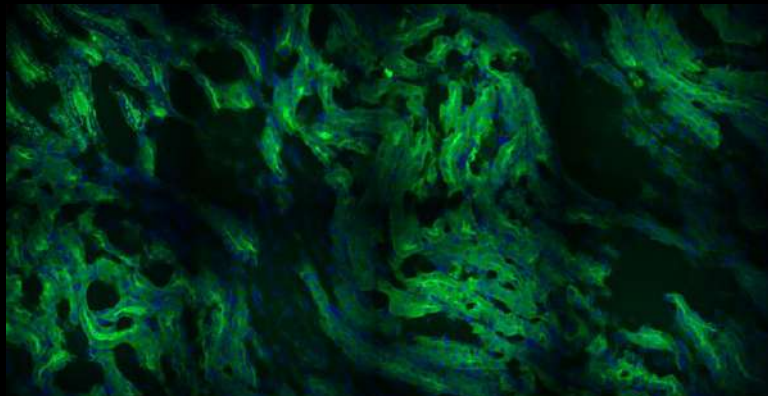
Standard Illumination

Borealis enhanced Illumination

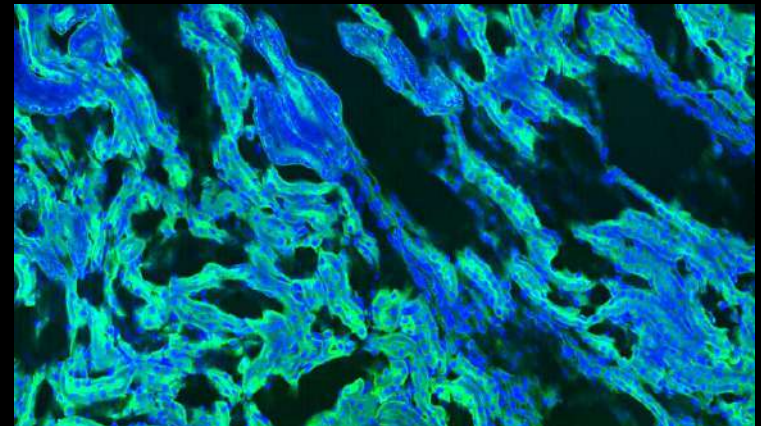


Borealis Perfect Illumination Delivery

Broad Spectrum Uniformity



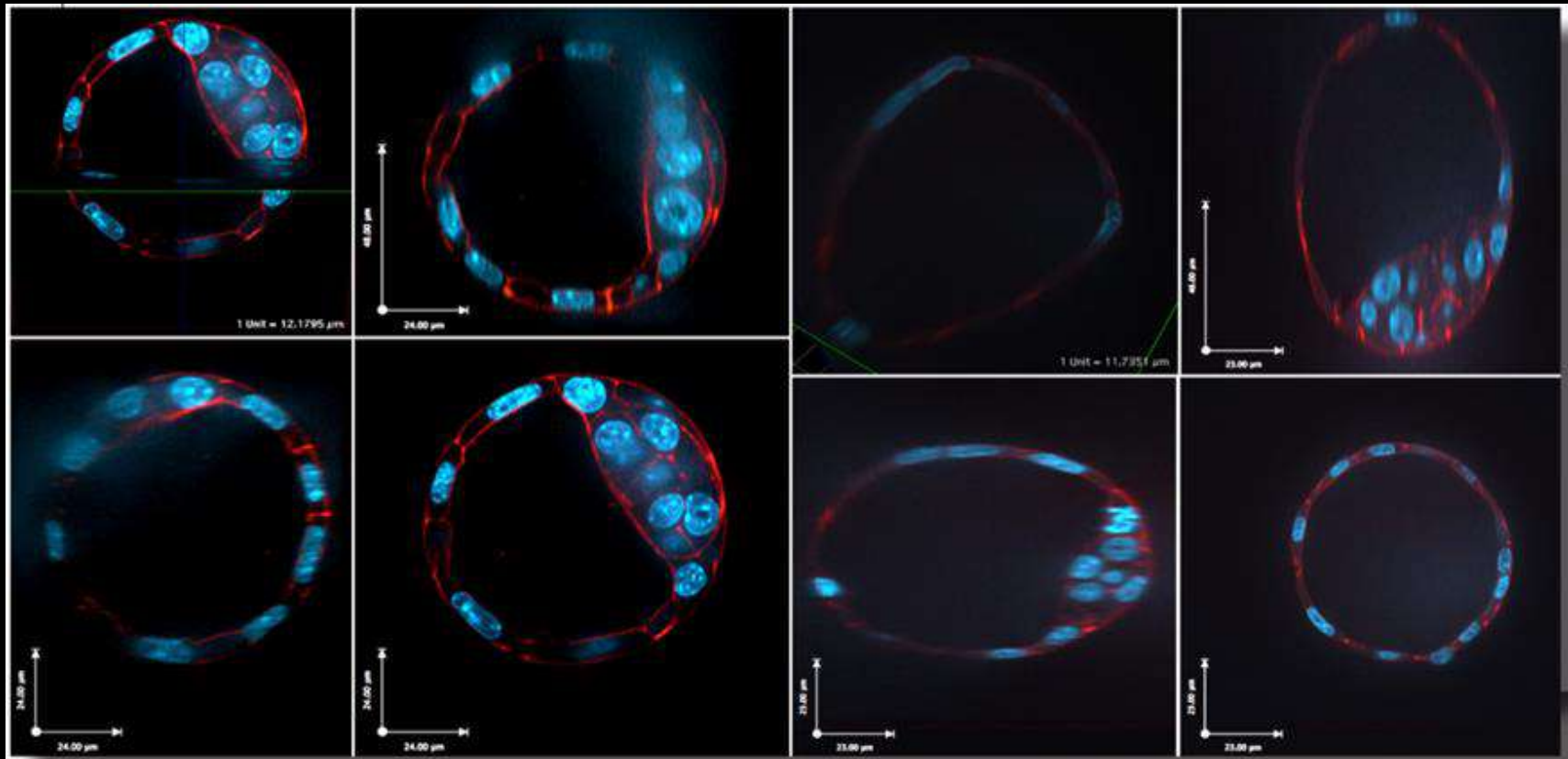
Standard CSUW1: exposure = 0.2s



Borealis Illumination: exposure = 0.2s

3x Signal - Higher contrast

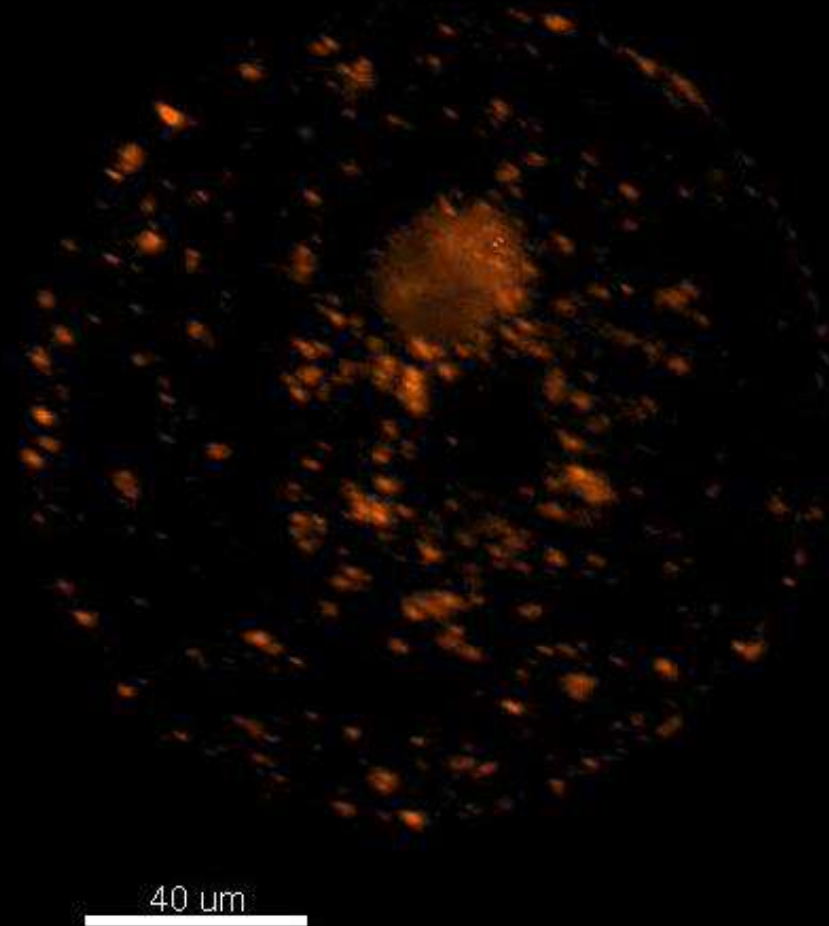
Minimizing Imaging Artefacts and Improving Image Quality



- With Borealis

- No Borealis

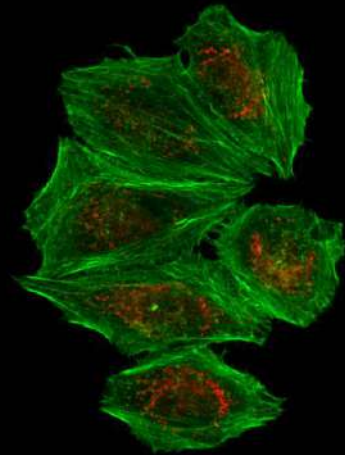
3D visualisation of a mouse egg stained for a plasma membrane protein. 600 focal planes over 80 microns



Curtesy of Katerina Dvorakova-Hortova, Laboratory of Reproduction, Institute of Biotechnology, CAS, v.v.i., BIOCEV, Czech Republic.

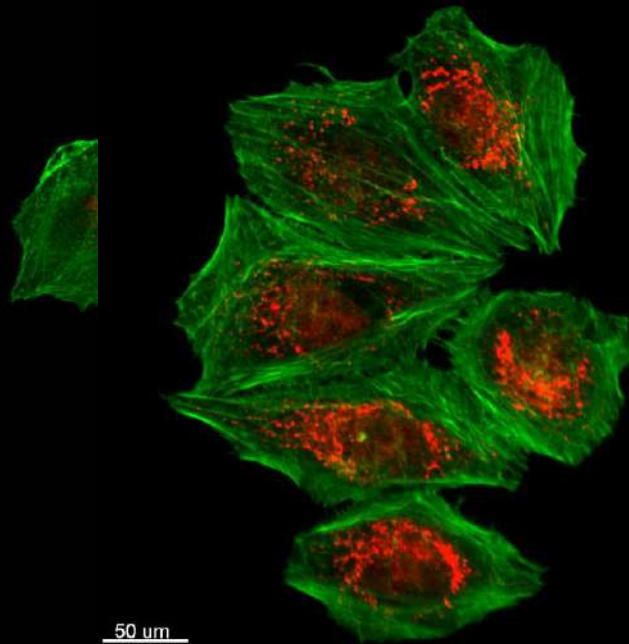
3 position zoom in camera port

1.0x



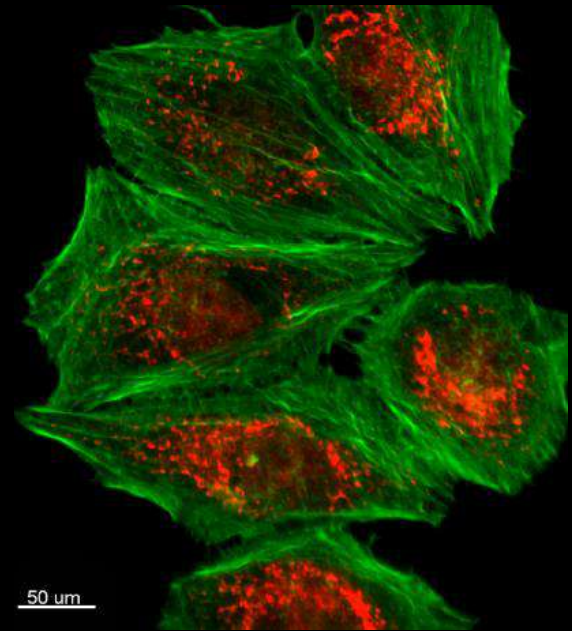
50 μ m

1.5x



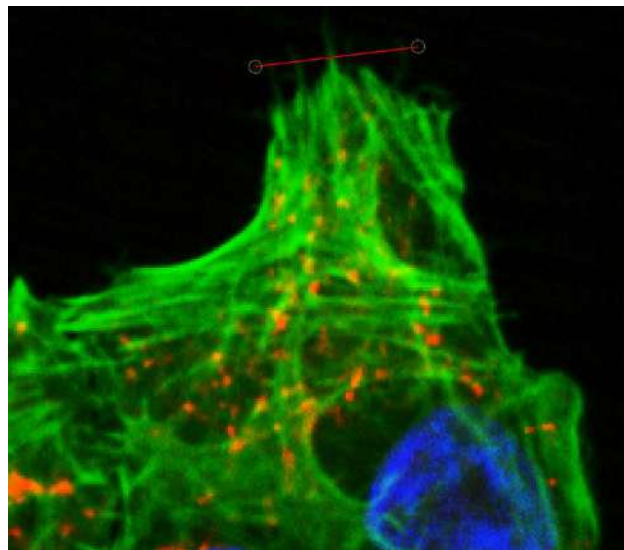
50 μ m

2.0x

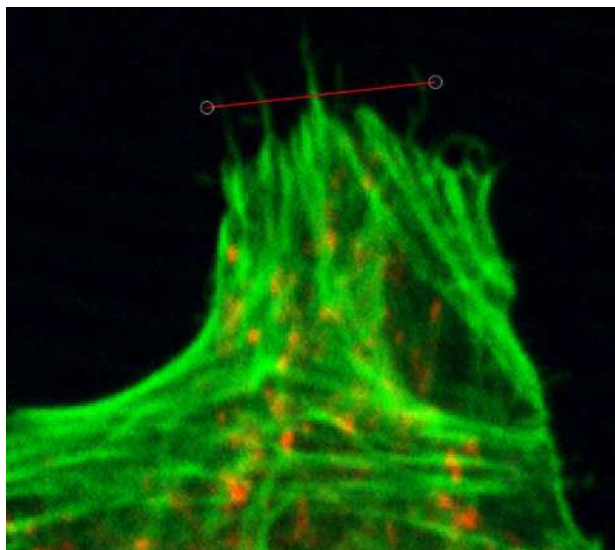
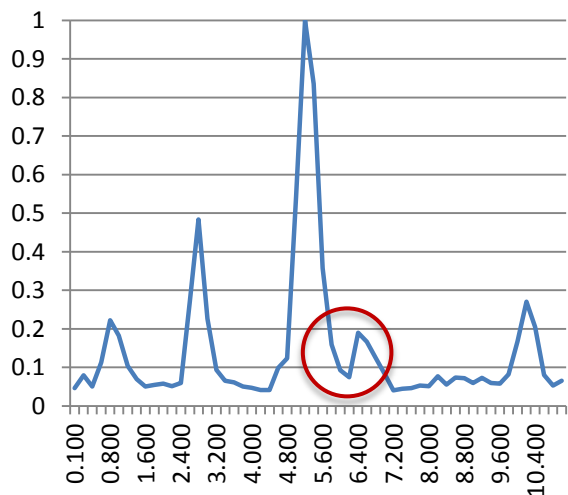


50 μ m

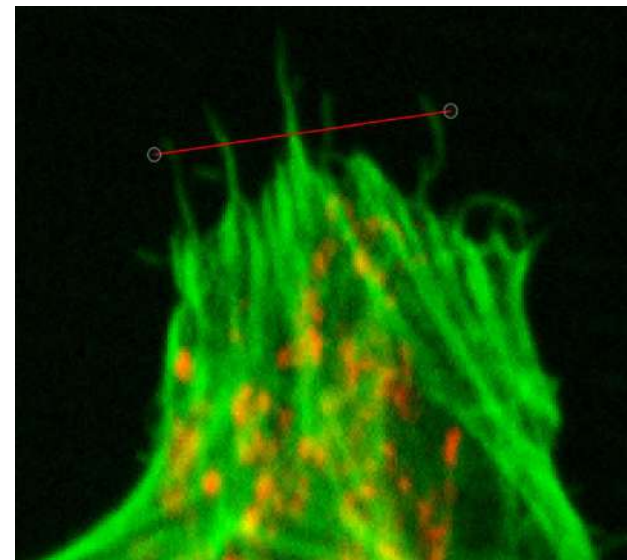
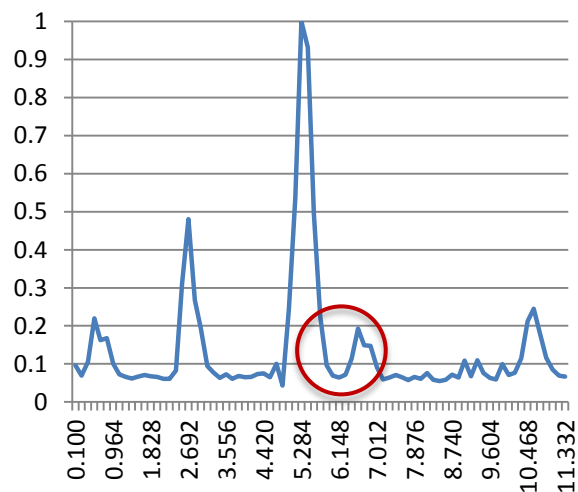
iXon Ultra 888. Actin filaments captured with 60x objective with varying additional camera magnification



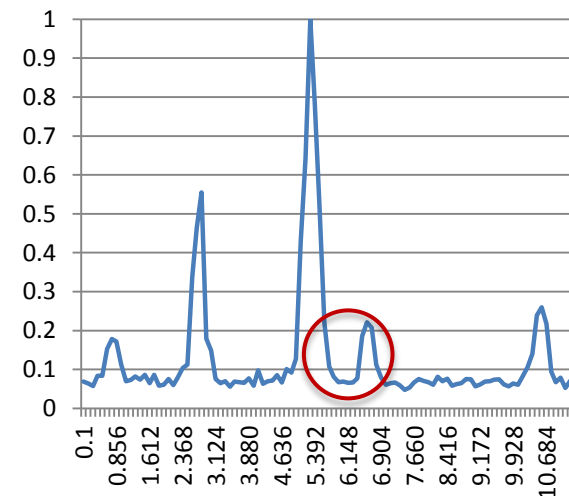
Actin 1.0x



Actin 1.5x

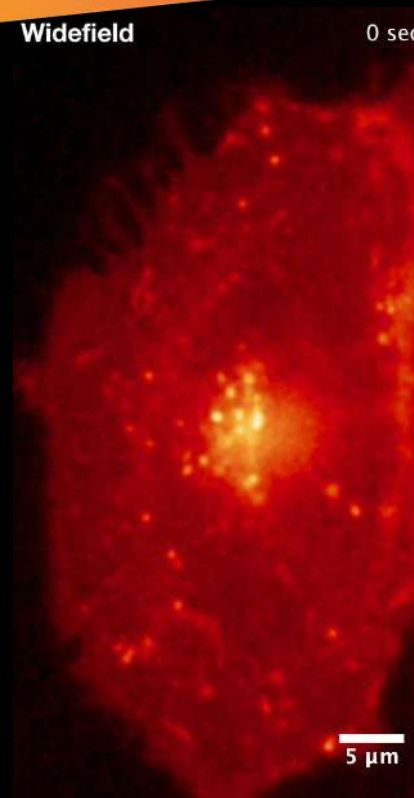
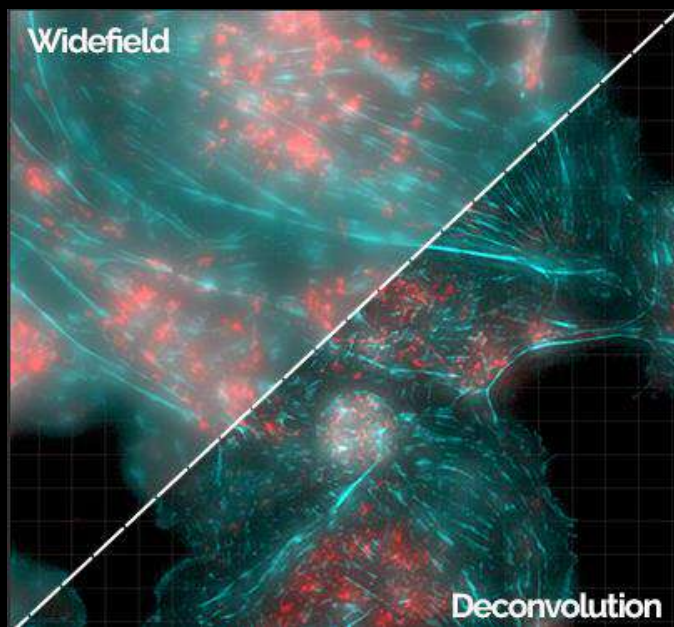
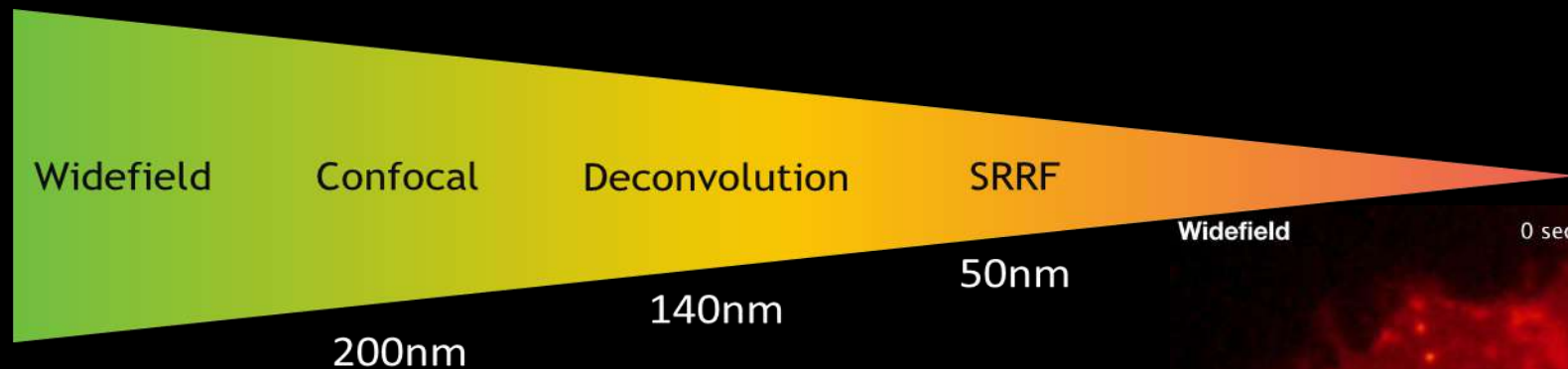


Actin 2.0x



Resolution

Multiple imaging techniques offer resolution down to 50nm (super-resolution)



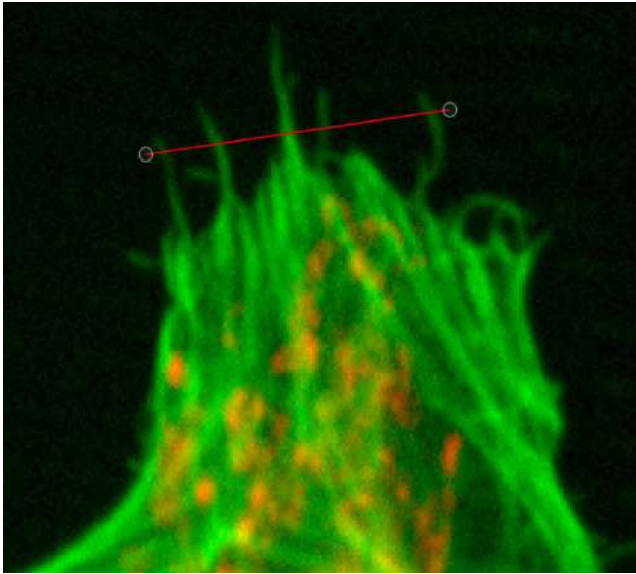
GPU accelerated deconvolution

The screenshot displays the ANDOR Fusion software interface, which is used for controlling microscopy acquisition and processing. The interface is divided into several panels:

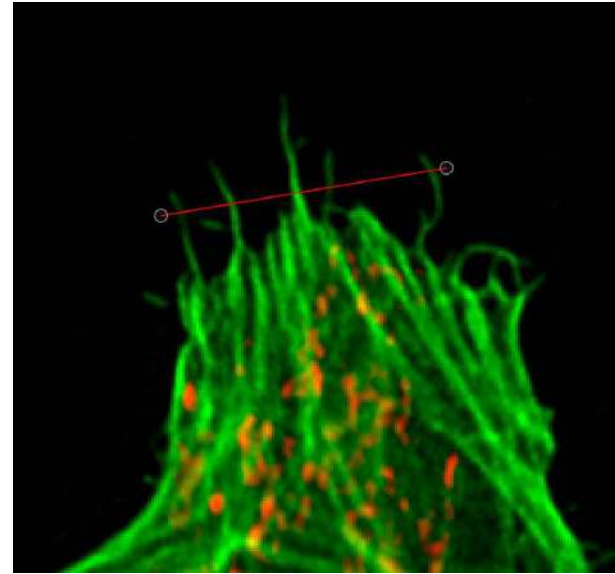
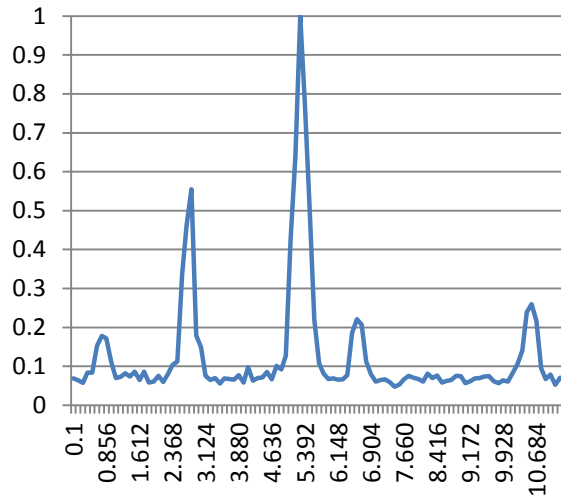
- Acquisition Control:** This panel on the left contains controls for live acquisition and snapping. It shows the active channel as 'RFP' in 'Confocal' mode. Parameters include Laser (488nm), EM Gain (0.0%), and Exposure Time (200 ms). Navigation controls and specimen management options are also present.
- Image Processing:** The right-hand panel is dedicated to image processing. It features a 'Deconvolving' button and a progress indicator. Under 'ADVANCED PROCESSING OPTIONS', various settings are visible:
 - Edge Artifact Reduction: All (XYZ)
 - Minimum Intensity Removal: ON
 - Denoising Filter Size: 0.7
 - Initial Denoising: ON
 - Denoising Frequency: 4
 - Normalisation: OFF
 - Number of Iterations: 24
 - Iteration Acceleration: OFF
- POINT SPREAD FUNCTION SETTINGS:** This section includes settings for the imaging modality (Spinning Disk Confocal), specimen refractive index (1.365), pinhole radius (40.0), magnification (100), objective lens NA (1.4), and immersion medium (Oil (1.52)).
- Current Calibration Information:** A bottom section showing details for the current channel, including microscope models like ALC-1111, X-1383, and CSU-666, and channel-specific parameters like 'Left' and '0 mm, 0 mm, 133.69 µm'.

The main window shows a green fluorescence image of a specimen. The status bar at the bottom indicates 'Multi' mode, 'ZOOM: Fit (45%)', and 'Pos' coordinates at 'x 1024 y 1024'.

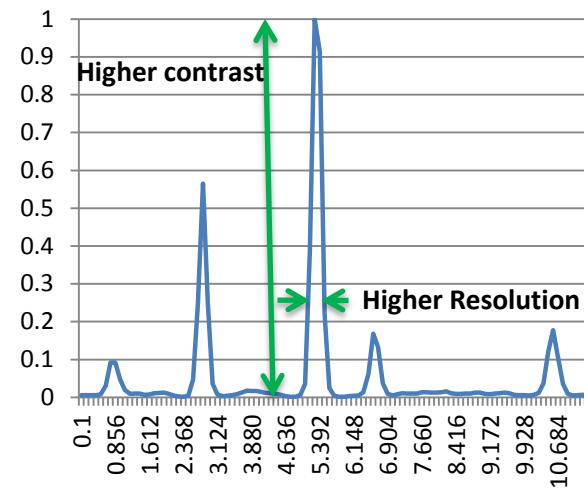
iXon Ultra 888. Actin filaments captured with 60x objective with 2.0x camera magnification. Deconvolution comparison.



Actin 2.0x

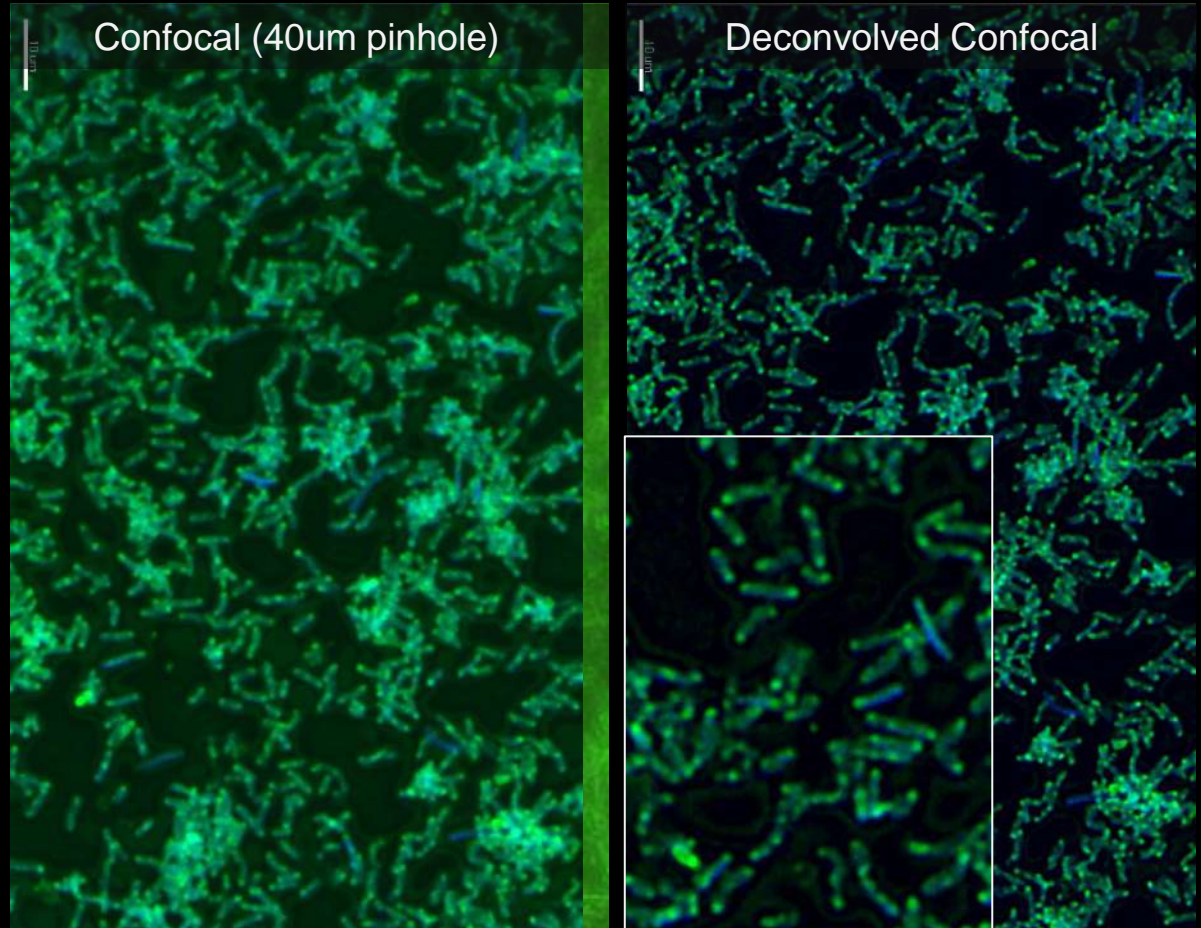


Actin 2.0x Deconvolved



Bacteria on biofilm

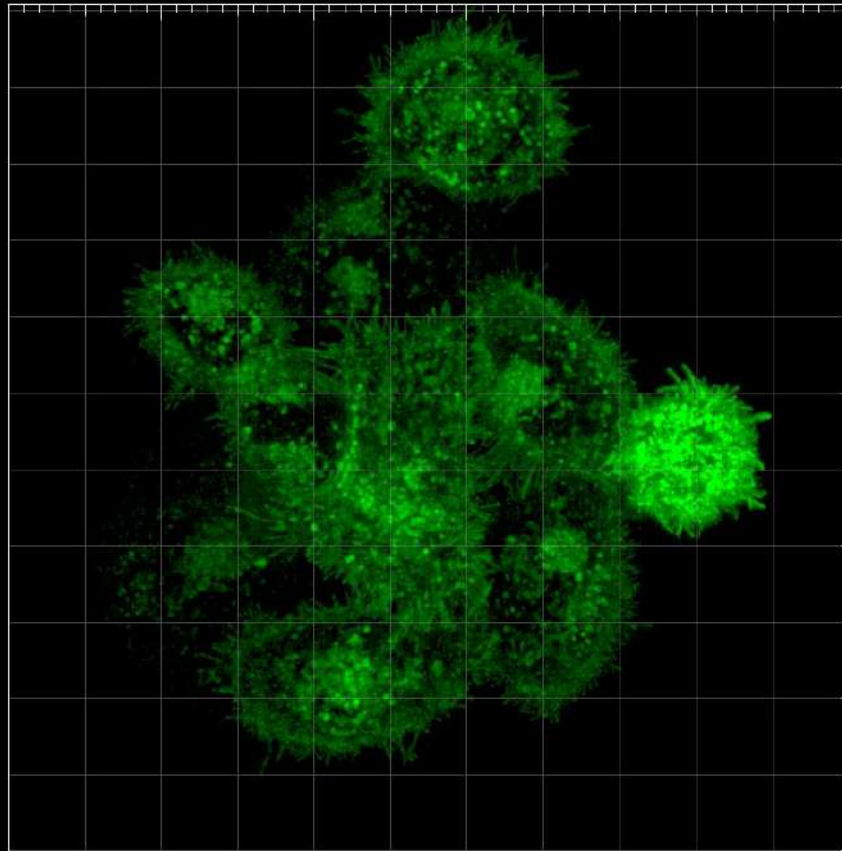
40x (0.75 NA) objective. Zyla 4.2



Dr Nigel Ternan & Dr Barry O'Hagan, University of Ulster

Confocal and Deconvolution

Hela cells expressing GFP fusion protein that is present in focal structures and vesicles. Deconvolution shows improved optical sectioning and enhancement in contrast. Images from same field with Dragonfly
– 288 sections , 0.1 μm Z step,

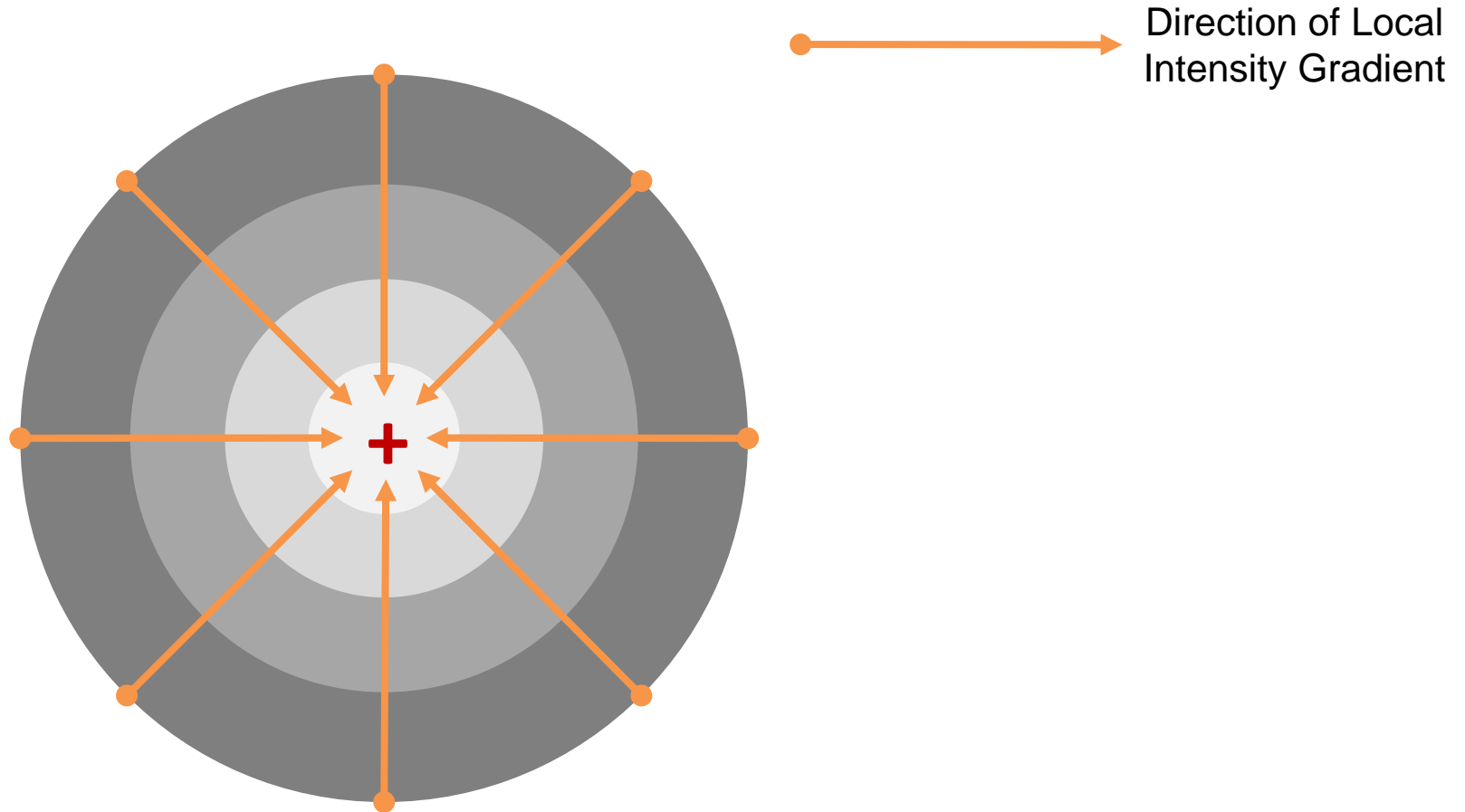


Super-resolution with SRRF

A living sample solution

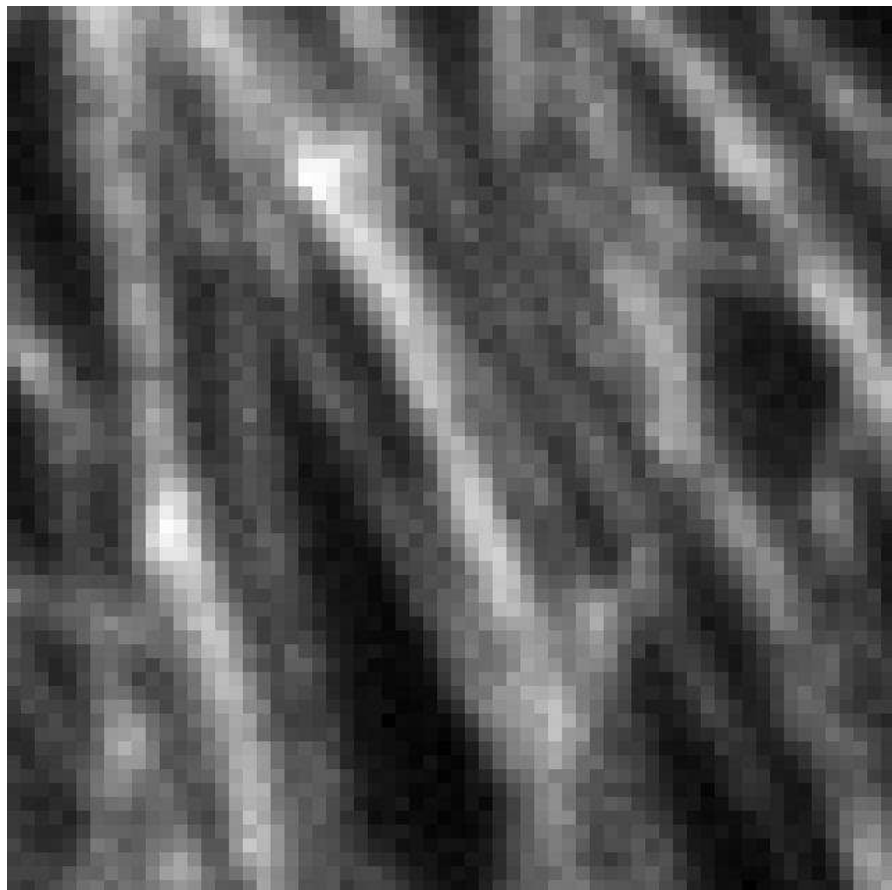
Combine with confocal

Super Resolution Radial Fluctuations

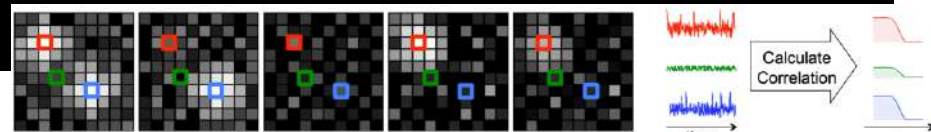
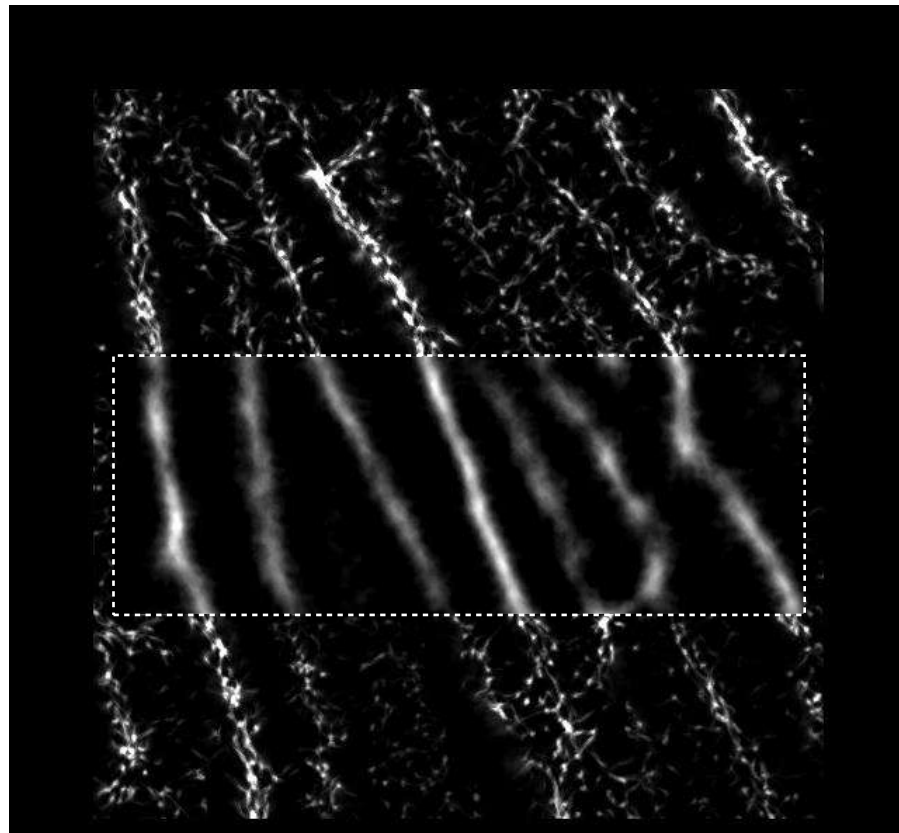


Fluorescent Marker is at the centre of intensity gradient field
aka **Point of maximum radial symmetry**

Super Resolution Radial Fluctuations

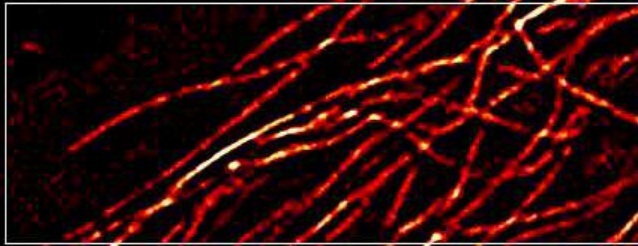
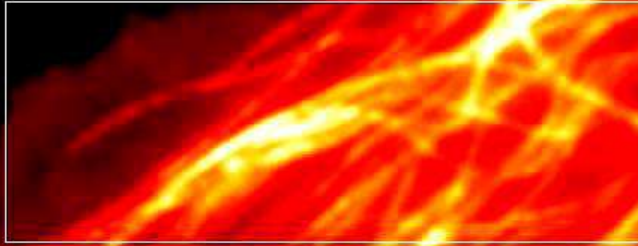


100 frames at 40mW/cm²



Live-cell, labeled with GFP

0 s



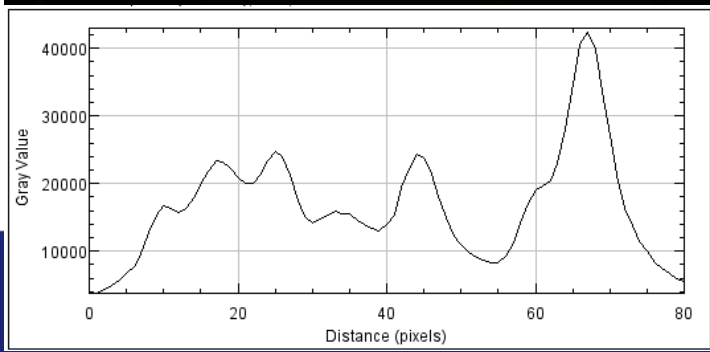
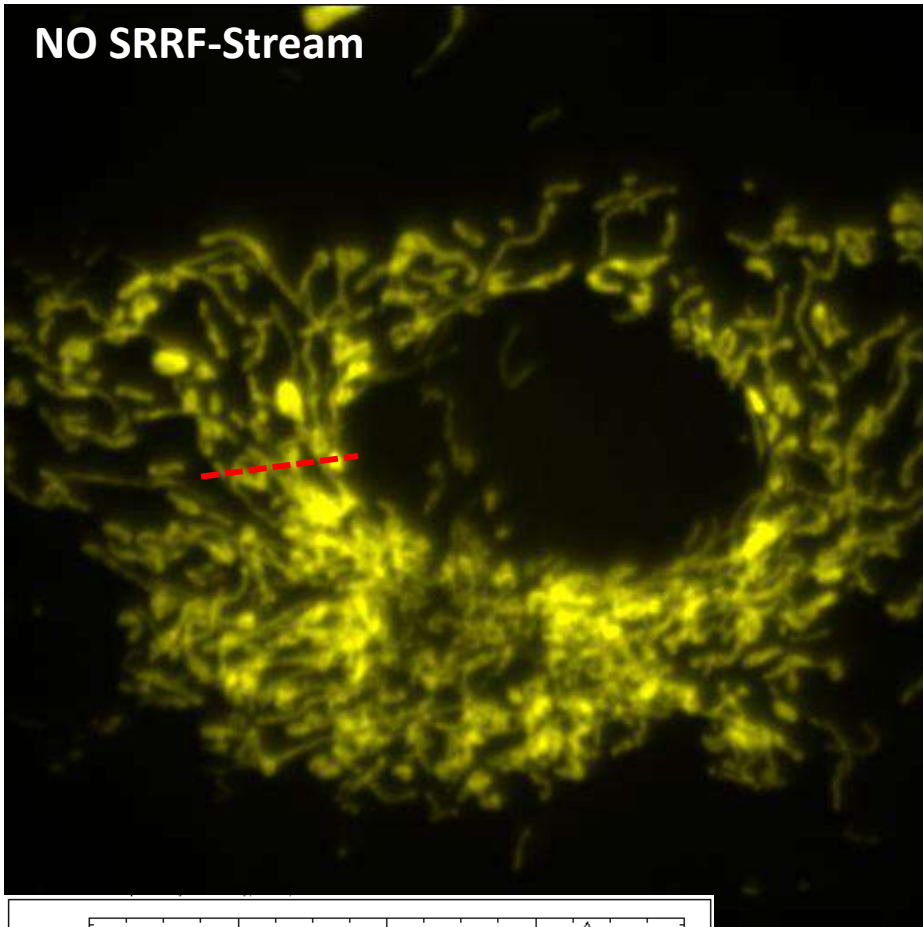
5 μ m

(8.5W/cm²) and no special optics

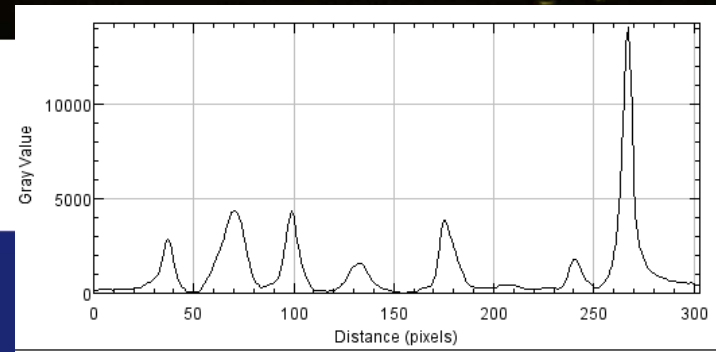
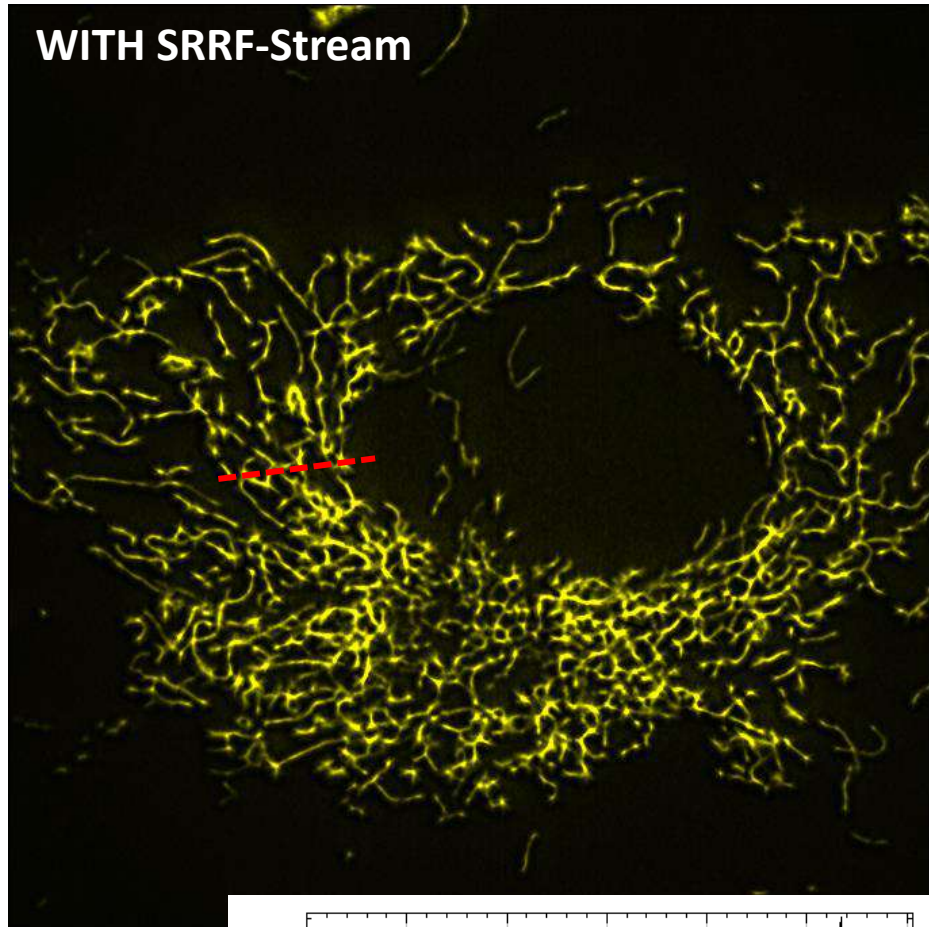
WideField SRRF – 560nm
Mitochondria

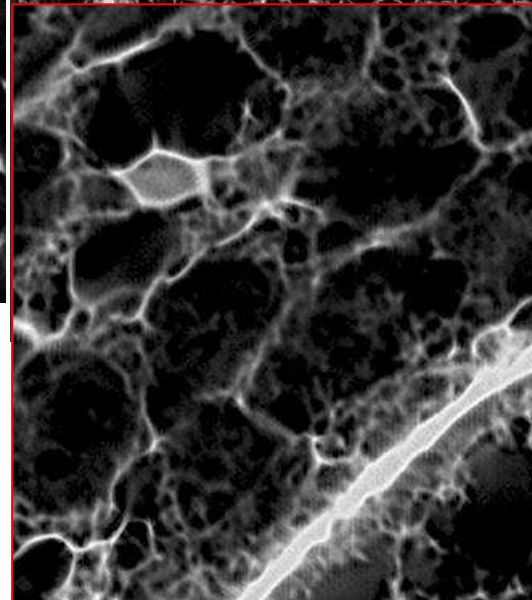
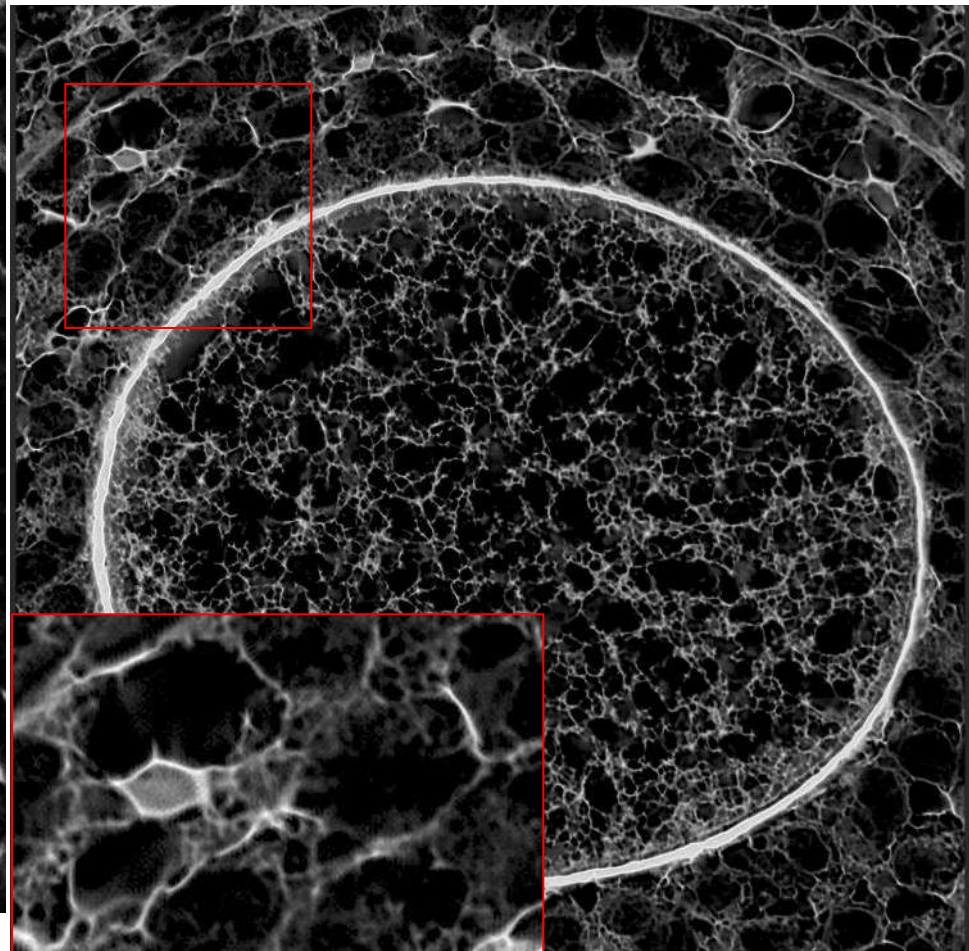
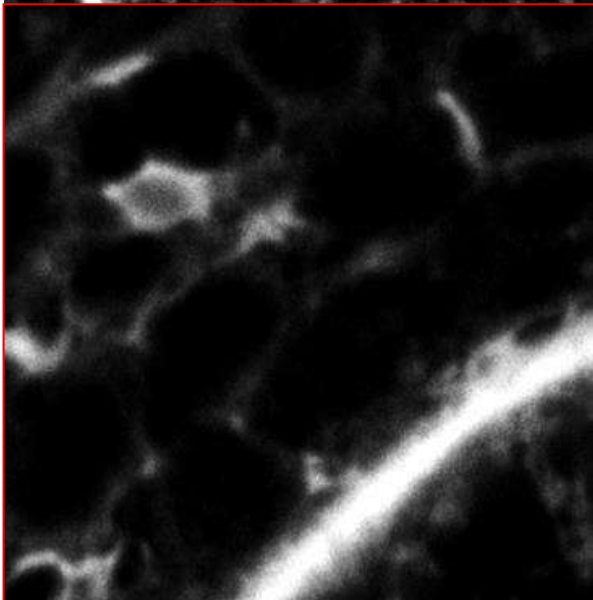
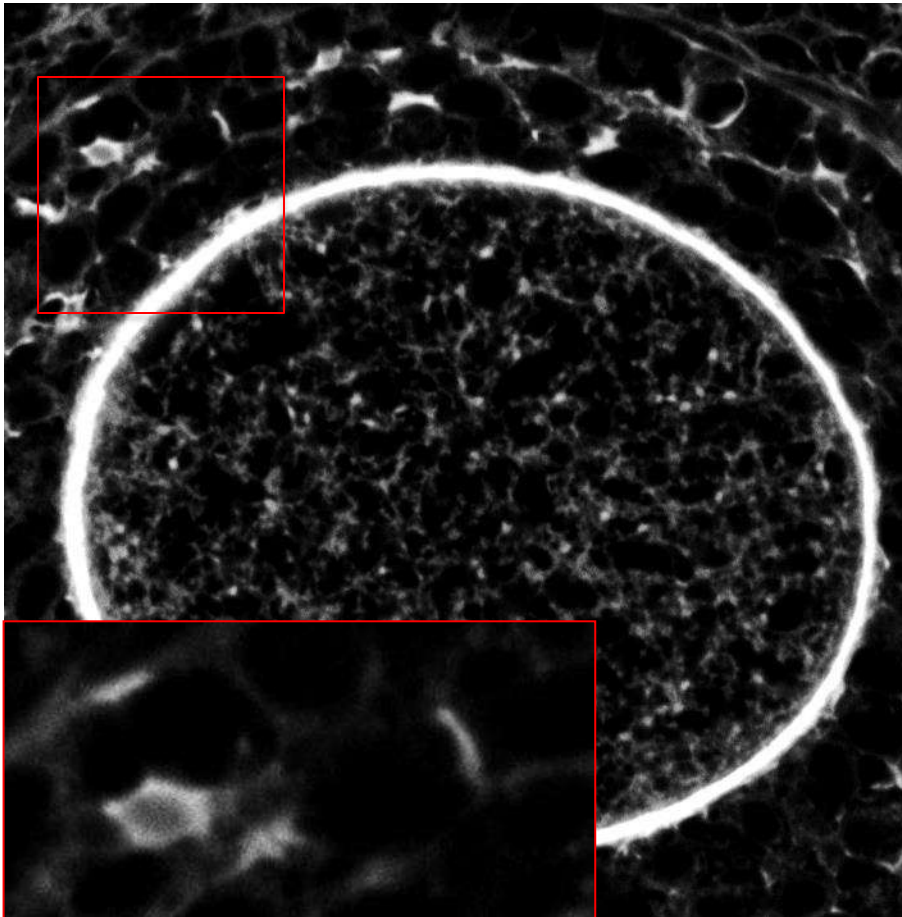
iXon Ultra 888
60x TIRF objective
2x camera magnifier
100 images – Average vs SRRF

NO SRRF-Stream



WITH SRRF-Stream





Ovary cross section
100 frames, 20 ms
Radial mag: 4X

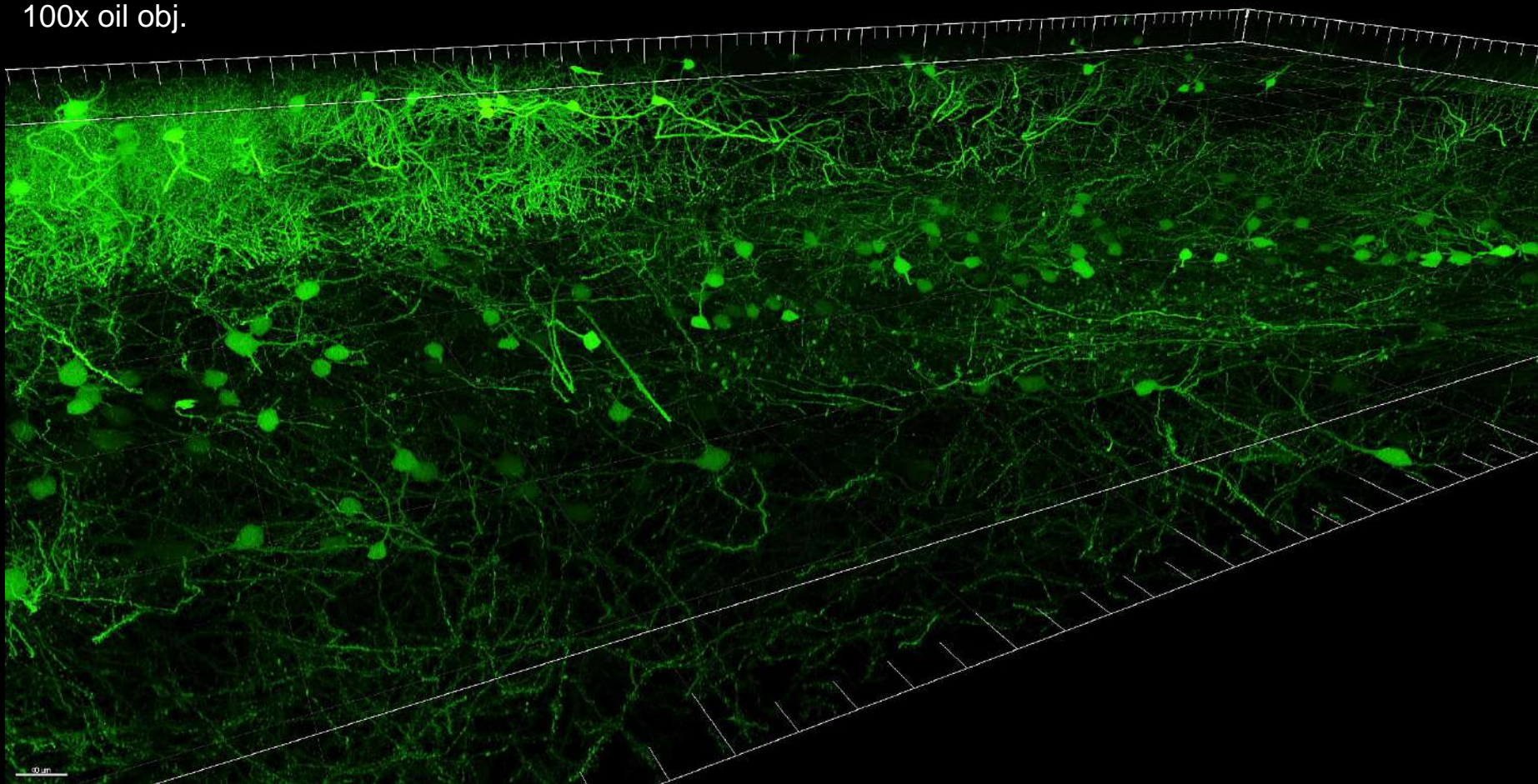
Confocal with SRRF – Brain Neuron

Confocal + SRRF

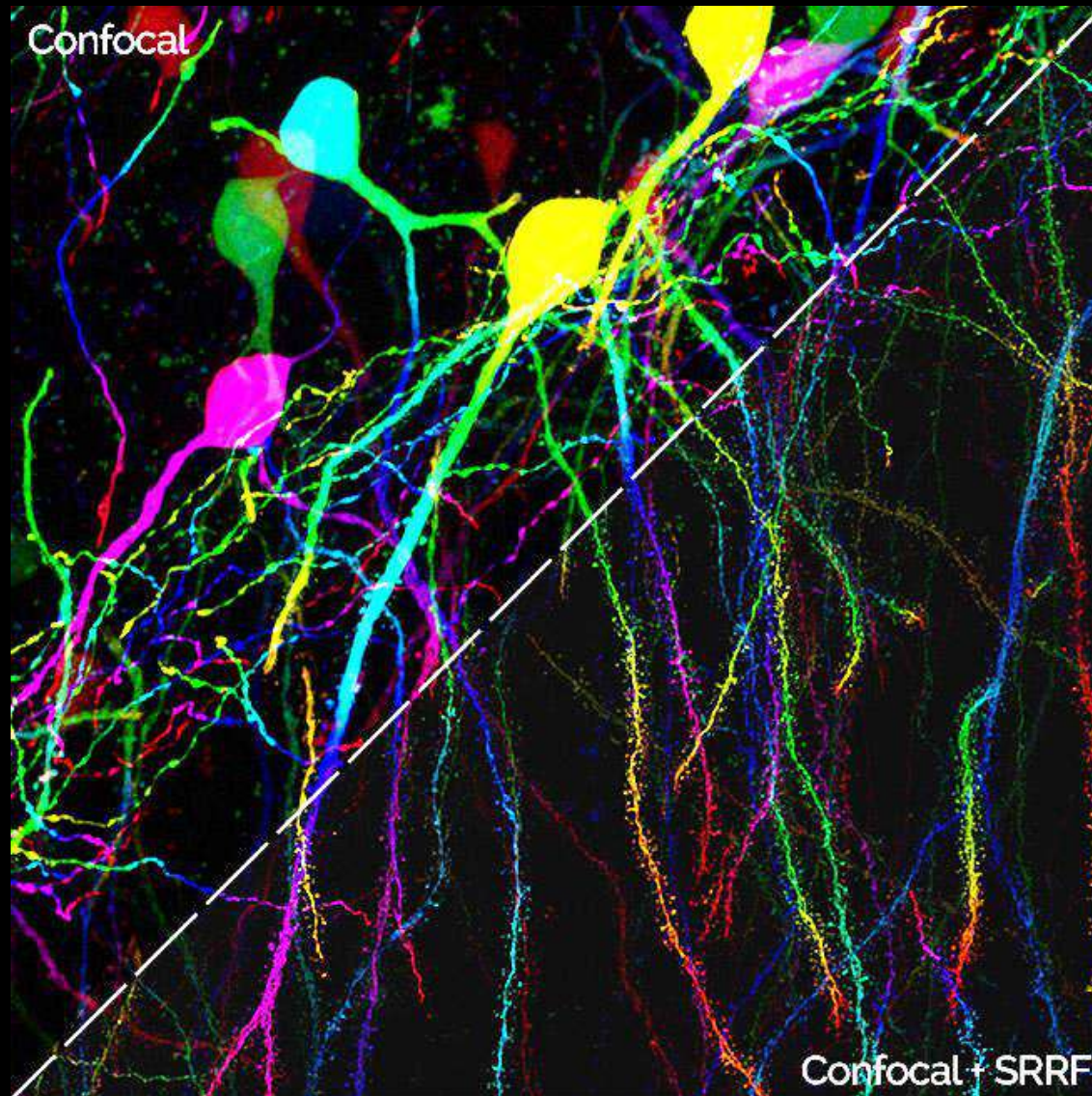
16x12 stitch

50um depth

100x oil obj.



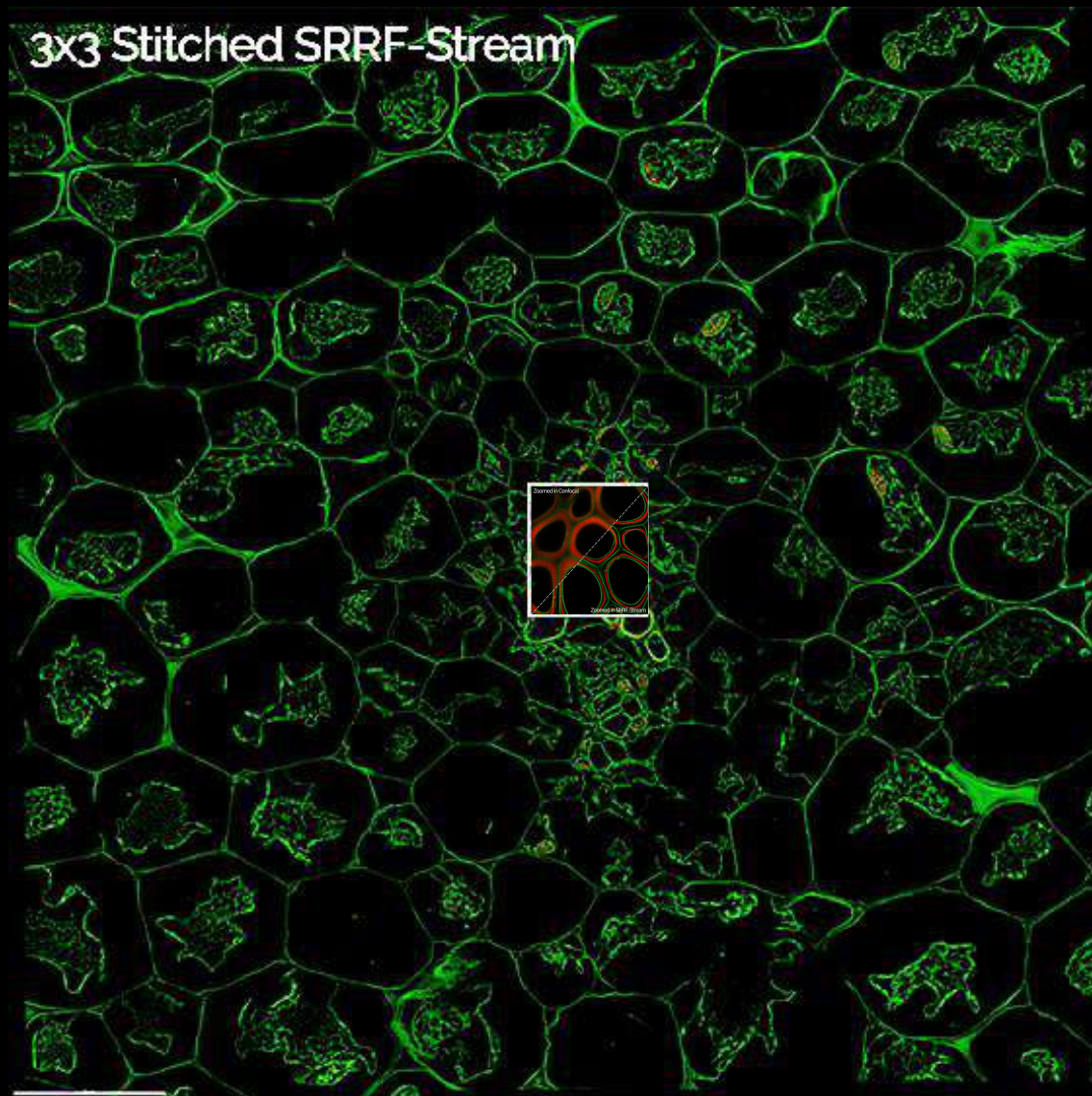
Confocal with SRRF – Brain Neuron



RapiClear
YFP labelled
neurons

Courtesy of
Sunjin Lab

From nanometer to millimeter...



From nanometer to millimeter...

